

[illegible]

Approved for use through 09/30/00. OMB 061-0032
Patent and Trademark Office: U.S. DEPARTMENT OF COMMERCE

+

(Only for new nonprovisional applications under 37 CFR 1.53(b))

First Named Inventor or Application Identifier

SU, et al.

Date	October 12, 1999
------	------------------

See MPEP chapter 600 concerning utility patent application contents.

Assistant Commissioner for Patents

Address to: Box Patent Application
Washington, D.C. 20231

- | | |
|---|---|
| <p>1. <input checked="" type="checkbox"/> Fee Transmittal Form</p> <p>2. <input checked="" type="checkbox"/> Specification [total pgs. 62]
(preferred arrangement set forth below)</p> <p>- Descriptive title of the Invention</p> <p>Access <input checked="" type="checkbox"/> References to Related Applications</p> <p>- Statement Regarding Fed Sponsored R&D</p> <p>- Reference to Microfiche Appendix</p> <p>- Background of the Invention</p> <p>- Brief Summary of the Invention</p> <p>- Brief Description of the Drawings (if filed)</p> <p>- Detailed Description</p> <p>- Claim(s)</p> <p>- Abstract of the Disclosure</p> <p>3. <input checked="" type="checkbox"/> Drawing(s):</p> <p>- Informal: Figs. 1-2 (2 sheets)</p> <p>- Formal: Figs. 1-2 (2 sheets)</p> <p>Total Sheets: 4</p> <p>4. <input checked="" type="checkbox"/> Oath or Declaration <u>Unexecuted (2 pp.)</u></p> <p>a. <input type="checkbox"/> Newly executed (original or copy)</p> <p>b. <input type="checkbox"/> Copy from a prior application (1.63(d))
(for con/div with Box 17 completed)</p> <p>i. <input type="checkbox"/> Deletion of Inventors
(signed statement attached deleting inventor(s) named in prior app.)</p> <p>5. <input checked="" type="checkbox"/> Incorporation by Reference (with Box 17)
The entire disclosure of the prior application, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein.</p> | <p>6. <input type="checkbox"/> Microfiche Computer Program (appendix)</p> <p>7. Nucleotide and/or Amino Acid Sequence Submission (if applicable, all necessary)</p> <p>a. <input type="checkbox"/> Computer Readable Copy</p> <p>b. <input checked="" type="checkbox"/> Paper Copy (identical to computer copy)
(6 pp.)</p> <p>c. <input type="checkbox"/> Statement verifying identity of above copies</p> |
| <p>Accompanying Application Parts</p> <p>8. <input checked="" type="checkbox"/> Assignment Papers (cover and document(s)):
Copy of: (a) Assignment, (b) Assignment Cover Sheet and (c) Notice of Recordation from parent application (SN 08/820,970) (7 pp.)</p> <p>9. <input type="checkbox"/> 37 CFR 3.73(b) Statement <input type="checkbox"/> Power of Attny</p> <p>10. <input type="checkbox"/> English Translation Document</p> <p>11. <input checked="" type="checkbox"/> Information Disclosure Statement/PTO-1449
<input checked="" type="checkbox"/> Copies of IDS Citations (AA-AG)</p> <p>12. <input checked="" type="checkbox"/> Preliminary Amendment (4 pp.)</p> <p>13. <input checked="" type="checkbox"/> Return Receipt Postcard (MPEP 503)
(should be specifically itemized)</p> <p>14. <input type="checkbox"/> Small Entity Statement(s)
<input type="checkbox"/> Statement filed in prior application, status still proper and desired</p> <p>15. <input type="checkbox"/> Certified copy of priority document(s)</p> <p>16. <input checked="" type="checkbox"/> Other: Request Under 37 CFR § 1.821(e)(1 pg.)</p> | |

17. If a CONTINUING APPLICATION, check appropriate box and supply the requisite information:

☐ Continuation ☐ Divisional ☒ Continuation-in-Part (CIP) of prior application Serial No.:
08/820,970, filed March 19, 1997

18. CORRESPONDENCE ADDRESS

☐ Customer Number or Bar Code Label or ☒ Correspondence address below

Name	Joseph J. Kenny., Agent for Applicants Human Genome Sciences, Inc.
------	---

Address	9410 Key West Avenue
---------	----------------------

City	Rockville
------	-----------

MD

20850

Country	US
---------	----

301-309-8504

301-309-8439

Joseph J. Kenny
Signature

Req. No. 43,710

10/12/99

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Attorney Docket No.: PF207P1

Date: October 12, 1999



Assistant Commissioner for Patents
Box PATENT APPLICATION
Washington, D.C. 20231

Sir:

Transmitted herewith for filing under 37 C.F.R. §1.53 is the following patent application:

Applicants: SU et al.

Title of Invention: Human Cytokine Polypeptide

Including: -Specification (57 pgs);
Claims (4 pgs); and
Abstract (1 pg).

Drawings: Figs. 1-2 (2 sheets)(informal)
Figs. 1-2 (2 sheets)(formal)

Sequence Listing: 6 pages

PATENT APPLICATION FEE VALUE

The filing fee of the subject application being filed concurrently herewith has been calculated, as follows:

TYPE	NO. FILE	LESS	EXTRA	EXTRA RATE	FEE
Total Claims	24	-20	4	\$18.00 each	72.00
Independent	3	-3	0	\$78.00 each	.00
Minimum Fee					760.00
Multiple Dependency Fee if applicable (\$260.00)					
Total Filing Fee					832.00

Priority of Provisional Application Serial No. 60/013,613, filed March 20, 1996, and U.S. Application Serial No. 08/820,970, filed March 19, 1997, is hereby claimed under 35 U.S.C. §120.

Please charge the required fee to Deposit Account No. 08-3425. In addition, the Commissioner is hereby authorized to charge payment for any additional filing fees required under 37 C.F.R. 1.16 or credit any overpayment to Deposit Account No. 08-3425. A duplicate of this paper is attached.

Respectfully submitted,

Dated: 10/12/99

Joseph J. Kenny
Joseph J. Kenny (Reg. No. 43,710)
Agent for Applicants

Human Genome Sciences, Inc.
9410 Key West Avenue
Rockville, MD 20850
Telephone: (301) 610-5800

MMW/JJK/lcc
Enclosures

09/416267 10/12/99

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of: SU et al.

Serial No.: To be assigned

Group Art Unit: To be assigned

Filed: HEREWITH

Examiner: To be assigned

Title: **Human Cytokine Polypeptide**

Atty. Docket No. PF270P1

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents
Washington, D.C. 20231

Sir:

Prior to examination, applicants hereby request that the following amendments and remarks be entered into the above-identified application. Please amend the application as follows.

In the Drawings

Please replace informal Figures 1 and 2 with formal Figures 1 and 2, attached to the application filed concurrently herewith.

In the Specification

Please amend the specification as follows.

Page 1, line 7, please insert the following paragraph:

-- This application claims benefit of priority to U.S. Application Serial No. 08/820,970, filed March 19, 1997 under 35 U.S.C. § 120, which claims priority to U.S. Provisional Application Serial No. 60/013,613, filed March 20, 1996 under 35 U.S.C. Section 119(e), and each of which is hereby incorporated by reference in its entirety. --; and

lines 15 and 16, please delete in their entirety.

Page 6, line 10, delete "g" and insert --μg--;

line 11, delete "l" and insert --μl--; and

0944667 104299

line 12, delete “g” and insert --μg--.

Page 24, line 19, after “residues about 203 to about” please delete “226” and replace therefor “223”.

Page 43, line 30, delete “g/kg” and insert --μg/kg--.

Page 44, line 3, delete “g/kg” and insert --μg/kg--.

Page 47, line 23, delete “g” and insert --μg--.

Page 51, line 23, delete “g” and insert --μg--;

line 24, delete “g” and insert --μg--;

line 26, delete “g” and insert --μg--;

line 27, delete “g” and insert --μg--;

line 28, delete “l” and insert --μl--;

line 29, delete “l” and insert --μl--; and

line 30, delete “l” and insert --μl--.

Page 52, line 18, delete “l” and insert --μl--;

line 21, after “4” insert --°--; and

line 30, delete “Ci” and insert --μCi--, both instances.

Page 55, line 30, after “65” insert --°--.

Page 56, line 2, after “60” insert --°--;

line 3, after “-70” insert --°--; and

line 16, after “37” insert --°--.

On page 19, line 15, please delete "12301 Park Lawn Drive, Rockville, MD 20852" and replace thereto --10801 University Boulevard, Manassas, VA 20110-2209--.

REMARKS

The specification has been amended to claim benefit of priority of the copending applications and to include the current address of the American Type Culture Collection (ATCC).

The amendment to the drawings is directed to a matter of form.

The amendments requesting that a "μ" be inserted in front of "g" or "l" in the specification find clear support in the provisional application, 60/013,613, to which the present application claims priority. In the process of converting the parent application, to which the present application claims priority, from a provisional application to a non-provisional application, a different word processing program was used. This word processing program failed to recognize the special character "μ" or "μ" leaving a blank space in the non-provisional document.

The following chart corresponds the location of the blank space in the non-provisional document with the position in the provisional application that evidences the existence of a "μ" or "μ" in applicants' priority document.

Amendment of the Present, Non-Provisional Application	Support in Provisional Application
Page 6, line 10, delete "g" and insert --μg--; line 11, delete "l" and insert --μl--; line 12, delete "g" and insert --μg--.	Page 6, line 7 Page 6, line 8 Page 6, line 10
Page 43, line 30, delete "g/kg" and insert --μg/kg--.	Page 42, line 12
Page 44, line 3, delete "g/kg" and insert --μg/kg--.	Page 42, line 15
Page 47, line 23, delete "g" and insert --μg--.	Page 45, line 17
Page 51, line 23, delete "g" and insert --μg--; line 24, delete "g" and insert --μg--; line 26, delete "g" and insert --μg--;	Page 49, line 7 Page 49, line 8 Page 49, line 11

line 27, delete "g" and insert --μg--; line 28, delete "I" and insert --μl--; line 29, delete "I" and insert --μl--; line 30, delete "I" and insert --μl--.	Page 49, line 12 Page 49, line 14 Page 49, line 15 Page 49, line 15
Page 52, line 18, delete "I" and insert --μl--; * line 21, after "4" insert --°--; and line 30, delete "Ci" and insert --μCi-- both instances.	Page 49, line 39 Page 50, line 4 Page 50, line 14
Page 55, line 30, after "65" insert --°--.	Page 53, line 6
Page 56, line 2, after "60" insert --°--; line 3, after "-70" insert --°--; and line 16, after "37" insert --°--.	Page 53, line 8 Page 53, line 9 Page 53, line 25

Thus, because the errors are typographical in nature and plain on their face to one of ordinary skill in the art, no new matter will be added by way of the amendments which add a "μ" in front of "g" or "I" or a "°" in front of "C."

No new matter has been added by way of amendment.

Entry of the above amendments and remarks is respectfully requested.

If there are any fees due in connection with the filing of this paper, please charge the fees to our Deposit Account No. 08-3425.

Respectfully submitted,

Dated: 10/12/99


Joseph J. Kenny (Reg No. 43,710)
Agent for Applicants

Human Genome Sciences, Inc.
9410 Key West Avenue
Rockville, MD 20850
Telephone: (301) 610-5800

MMW/lcc

HUMAN CYTOKINE POLYPEPTIDE

This invention relates, in part, to newly identified polynucleotides and polypeptides; variants and derivatives of the polynucleotides and polypeptides; processes for making the polynucleotides and the polypeptides, and their variants and derivatives; agonists and antagonists of the polypeptides; and uses of the polynucleotides, polypeptides, variants, derivatives, agonists and antagonists. In particular, in these and in other regards, the invention relates to polynucleotides and polypeptides of human cytokine polypeptide.

This application claims benefit of 35 U.S.C. 120 based on copending U.S. Provisional Application Serial No.: 60/013613, filed on March 20, 1996.

BACKGROUND OF THE INVENTION

The human cytokine polypeptide of the present invention has a high degree of similarity to the *Drosophila* twisted gastrulation gene and also has homology to human connective tissue growth factor.

The twisted gastrulation gene (TSG) is one of seven known zygotic genes that specify the fate of dorsal cells in *Drosophila* embryos. Mutations in these genes cause at least some of the cells on the dorsal half of the embryo to adopt more ventral cell fates leading to the proposal that most of these genes participate in establishing, maintaining or modulating a gradient of a single signaling molecule decapentaplegic. The effects of TSG mutations on the development of cuticle elements, expression of a region specific enhancer trap and patterns of mitotic domains have been examined. Mutations of TSG only affect the fate of a narrow strip of dorsal midline cells and do not affect dorsal ectoderm cells. However, the pattern of TSG expression is not coincident with the territories affected by TSG mutations. Structural analysis of the TSG gene reveals features of a secreted protein suggesting an extra cellular site of action. The TSG protein bears a weak resemblance to human connective tissue

growth factor (CTGF), a TGF- β -induced protein. The dorsal midline cell fate is specified by the combination of both a TSG and a decapentaplegic signal to which the dorsal midline cells are uniquely competent to respond. Mason, E. D. et al., Genes Dev., 8:1489-1501 (1994).

5 Accordingly, there is a need in the art for identification and characterization of proteins that influence biological activity, both normally and in disease states. In particular, there is a need to isolate and characterize human cytokine polypeptides akin to the TSG gene found in *Drosophila* which may be employed, therefore, for preventing, ameliorating or correcting dysfunctions or disease or augmenting positive
10 natural actions of such genes.

SUMMARY OF THE INVENTION

Toward these ends, and others, it is an object of the present invention to provide polypeptides, inter alia, that have been identified as novel human cytokine
15 polypeptide by homology between the amino acid sequence set out in Figure 1 (SEQ ID NO:2) and known amino acid sequences of other proteins such as the *Drosophila* twisted gastrulation gene (TSG) (SEQ ID NO:9).

It is a further object of the invention, moreover, to provide polynucleotides that encode human cytokine polypeptide, particularly polynucleotides that encode
20 the polypeptide herein designated human cytokine polypeptide.

In a particularly preferred embodiment of this aspect of the invention the polynucleotide comprises the region encoding human cytokine polypeptide in the sequence set out in Figure 1 (SEQ ID NO:2).

In accordance with this aspect of the present invention there is provided an
25 isolated nucleic acid molecule encoding a mature polypeptide expressed by the human cDNA contained in the deposited clone as hereinafter described.

In accordance with this aspect of the invention there are provided isolated nucleic acid molecules encoding human cytokine polypeptide, including mRNAs, cDNAs, genomic DNAs and, in further embodiments of this aspect of the invention,
30 biologically, diagnostically, clinically or therapeutically useful variants, analogs or

derivatives thereof, or fragments thereof, including fragments of the variants, analogs and derivatives.

Among the particularly preferred embodiments of this aspect of the invention are naturally occurring allelic variants of human cytokine polypeptide.

5 It also is an object of the invention to provide human cytokine polypeptides, particularly human cytokine polypeptide, that stimulates cell proliferation and/or differentiation and may be employed to treat and/or prevent restenosis and inflammation.

10 In accordance with this aspect of the invention there are provided novel polypeptides of human origin referred to herein as human cytokine polypeptide as well as biologically, diagnostically or therapeutically useful fragments, variants and derivatives thereof, variants and derivatives of the fragments, and analogs of the foregoing.

15 Among the particularly preferred embodiments of this aspect of the invention are variants of human cytokine polypeptide encoded by naturally occurring alleles of the human cytokine polypeptide gene.

20 It is another object of the invention to provide a process for producing the aforementioned polypeptides, polypeptide fragments, variants and derivatives, fragments of the variants and derivatives, and analogs of the foregoing. In a preferred embodiment of this aspect of the invention there are provided methods for producing the aforementioned human cytokine polypeptide comprising culturing host cells having expressibly incorporated therein an exogenously-derived human cytokine polypeptide-encoding polynucleotide under conditions for expression of human cytokine polypeptide in the host and then recovering the expressed polypeptide.

25 In accordance with another object the invention there are provided products, compositions, processes and methods that utilize the aforementioned polypeptides and polynucleotides for research, biological, clinical and therapeutic purposes, inter alia.

30 In accordance with certain preferred embodiments of this aspect of the invention, there are provided products, compositions and methods, inter alia, for,

among other things: assessing human cytokine polypeptide expression in cells by determining human cytokine polypeptide or human cytokine polypeptide-encoding mRNA; assaying genetic variation and aberrations, such as defects, in human cytokine polypeptide genes; and administering a human cytokine polypeptide or
5 polynucleotide to an organism to augment human cytokine polypeptide function or remediate human cytokine polypeptide dysfunction.

In accordance with certain preferred embodiments of this and other aspects of the invention there are provided probes that hybridize to human cytokine polypeptide sequences.

10 In certain additional preferred embodiments of this aspect of the invention there are provided antibodies against human cytokine polypeptides. In certain particularly preferred embodiments in this regard, the antibodies are highly selective for human cytokine polypeptide.

In accordance with another aspect of the present invention, there are provided
15 human cytokine polypeptide agonists. Among preferred agonists are molecules that mimic human cytokine polypeptide, that bind to human cytokine polypeptide-binding molecules or receptor molecules, and that elicit or augment human cytokine polypeptide-induced responses. Also among preferred agonists are molecules that interact with human cytokine polypeptide or human cytokine polypeptides, or with
20 other modulators of human cytokine polypeptide activities, and thereby potentiate or augment an effect of human cytokine polypeptide or more than one effect of human cytokine polypeptide.

In accordance with yet another aspect of the present invention, there are provided human cytokine polypeptide antagonists. Among preferred antagonists are
25 those which mimic human cytokine polypeptide so as to bind to human cytokine polypeptide receptor or binding molecules but not elicit a human cytokine polypeptide-induced response or more than one human cytokine polypeptide-induced response. Also among preferred antagonists are molecules that bind to or interact with human cytokine polypeptide so as to inhibit an effect of human cytokine

polypeptide or more than one effect of human cytokine polypeptide or which prevent expression of human cytokine polypeptide.

The antagonists may be used to inhibit the action of human cytokine polypeptides. They may be used, for instance, to inhibit abnormal cellular proliferation, for example, neoplasia.

In a further aspect of the invention there are provided compositions comprising a human cytokine polypeptide polynucleotide or a human cytokine polypeptide for administration to cells in vitro, to cells ex vivo and to cells in vivo, or to a multicellular organism. In certain particularly preferred embodiments of this aspect of the invention, the compositions comprise a human cytokine polypeptide polynucleotide for expression of a human cytokine polypeptide in a host organism for treatment of disease. Particularly preferred in this regard is expression in a human patient for treatment of a dysfunction associated with aberrant endogenous activity of human cytokine polypeptide.

Other objects, features, advantages and aspects of the present invention will become apparent to those of skill from the following description. It should be understood, however, that the following description and the specific examples, while indicating preferred embodiments of the invention, are given by way of illustration only. Various changes and modifications within the spirit and scope of the disclosed invention will become readily apparent to those skilled in the art from reading the following description and from reading the other parts of the present disclosure.

BRIEF DESCRIPTION OF THE DRAWINGS

The following drawings depict certain embodiments of the invention. They are illustrative only and do not limit the invention otherwise disclosed herein.

Figure 1 shows the nucleotide and deduced amino acid sequence of human cytokine polypeptide (SEQ ID NO:1 and 2).

Figure 2 shows the regions of similarity between amino acid sequences of human cytokine polypeptide (SEQ ID NO:2) and TSG polypeptide (SEQ ID NO:9).

GLOSSARY

The following illustrative explanations are provided to facilitate understanding of certain terms used frequently herein, particularly in the examples. The explanations are provided as a convenience and are not limitative of the invention.

5 DIGESTION of DNA refers to catalytic cleavage of the DNA with a restriction enzyme that acts only at certain sequences in the DNA. The various restriction enzymes referred to herein are commercially available and their reaction conditions, cofactors and other requirements for use are known and routine to the skilled artisan.

10 For analytical purposes, typically, 1 g of plasmid or DNA fragment is digested with about 2 units of enzyme in about 20 l of reaction buffer. For the purpose of isolating DNA fragments for plasmid construction, typically 5 to 50 g of DNA are digested with 20 to 250 units of enzyme in proportionately larger volumes.

15 Appropriate buffers and substrate amounts for particular restriction enzymes are described in standard laboratory manuals, such as those referenced below, and they are specified by commercial suppliers.

20 Incubation times of about 1 hour at 37 C are ordinarily used, but conditions may vary in accordance with standard procedures, the supplier's instructions and the particulars of the reaction. After digestion, reactions may be analyzed, and fragments may be purified by electrophoresis through an agarose or polyacrylamide gel, using well known methods that are routine for those skilled in the art.

25 GENETIC ELEMENT generally means a polynucleotide comprising a region that encodes a polypeptide or a region that regulates transcription or translation or other processes important to expression of the polypeptide in a host cell, or a polynucleotide comprising both a region that encodes a polypeptide and a region operably linked thereto that regulates expression.

30 Genetic elements may be comprised within a vector that replicates as an episomal element; that is, as a molecule physically independent of the host cell genome. They may be comprised within mini-chromosomes, such as those that arise during amplification of transfected DNA by methotrexate selection in eukaryotic cells.

Genetic elements also may be comprised within a host cell genome; not in their natural state but, rather, following manipulation such as isolation, cloning and introduction into a host cell in the form of purified DNA or in a vector, among others.

ISOLATED means altered "by the hand of man" from its natural state; i.e.,
5 that, if it occurs in nature, it has been changed or removed from its original environment, or both.

For example, a naturally occurring polynucleotide or a polypeptide naturally present in a living animal in its natural state is not "isolated," but the same polynucleotide or polypeptide separated from the coexisting materials of its natural
10 state is "isolated", as the term is employed herein. For example, with respect to polynucleotides, the term isolated means that it is separated from the chromosome and cell in which it naturally occurs.

As part of or following isolation, such polynucleotides can be joined to other polynucleotides, such as DNAs, for mutagenesis, to form fusion proteins, and for
15 propagation or expression in a host, for instance. The isolated polynucleotides, alone or joined to other polynucleotides such as vectors, can be introduced into host cells, in culture or in whole organisms. Introduced into host cells in culture or in whole organisms, such DNAs still would be isolated, as the term is used herein, because they would not be in their naturally occurring form or environment. Similarly, the
20 polynucleotides and polypeptides may occur in a composition, such as a media formulations, solutions for introduction of polynucleotides or polypeptides, for example, into cells, compositions or solutions for chemical or enzymatic reactions, for instance, which are not naturally occurring compositions, and, therein remain isolated polynucleotides or polypeptides within the meaning of that term as it is employed
25 herein.

LIGATION refers to the process of forming phosphodiester bonds between two or more polynucleotides, which most often are double stranded DNAs. Techniques for ligation are well known to the art and protocols for ligation are described in standard laboratory manuals and references, such as, for instance,
30 Sambrook et al., MOLECULAR CLONING, A LABORATORY MANUAL, 2nd

Ed.; Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (1989) and Maniatis et al., pg. 146, as cited below.

OLIGONUCLEOTIDE(S) refers to relatively short polynucleotides. Often the term refers to single-stranded deoxyribonucleotides, but it can refer as well to
5 single-or double-stranded ribonucleotides, RNA:DNA hybrids and double-stranded DNAs, among others.

Oligonucleotides, such as single-stranded DNA probe oligonucleotides, often are synthesized by chemical methods, such as those implemented on automated oligonucleotide synthesizers. However, oligonucleotides can be made by a variety of
10 other methods, including in vitro recombinant DNA-mediated techniques and by expression of DNAs in cells and organisms.

Initially, chemically synthesized DNAs typically are obtained without a 5' phosphate. The 5' ends of such oligonucleotides are not substrates for phosphodiester bond formation by ligation reactions that employ DNA ligases
15 typically used to form recombinant DNA molecules. Where ligation of such oligonucleotides is desired, a phosphate can be added by standard techniques, such as those that employ a kinase and ATP.

The 3' end of a chemically synthesized oligonucleotide generally has a free hydroxyl group and, in the presence of a ligase, such as T4 DNA ligase, readily will
20 form a phosphodiester bond with a 5' phosphate of another polynucleotide, such as another oligonucleotide. As is well known, this reaction can be prevented selectively, where desired, by removing the 5' phosphates of the other polynucleotide(s) prior to ligation.

PLASMIDS generally are designated herein by a lower case p preceded and/or
25 followed by capital letters and/or numbers, in accordance with standard naming conventions that are familiar to those of skill in the art.

Starting plasmids disclosed herein are either commercially available, publicly available on an unrestricted basis, or can be constructed from available plasmids by routine application of well known, published procedures. Many plasmids and other
30 cloning and expression vectors that can be used in accordance with the present

invention are well known and readily available to those of skill in the art. Moreover, those of skill readily may construct any number of other plasmids suitable for use in the invention. The properties, construction and use of such plasmids, as well as other vectors, in the present invention will be readily apparent to those of skill from the present disclosure.

POLYNUCLEOTIDE(S) generally refers to any polyribonucleotide or polydeoxribonucleotide, which may be unmodified RNA or DNA or modified RNA or DNA. Thus, for instance, polynucleotides as used herein refers to, among others, single-and double-stranded DNA, DNA that is a mixture of single-and double-stranded regions, single- and double-stranded RNA, and RNA that is mixture of single- and double-stranded regions, hybrid molecules comprising DNA and RNA that may be single-stranded or, more typically, double-stranded or a mixture of single- and double-stranded regions. In addition, polynucleotide as used herein refers to triple-stranded regions comprising RNA or DNA or both RNA and DNA. The strands in such regions may be from the same molecule or from different molecules. The regions may include all of one or more of the molecules, but more typically involve only a region of some of the molecules. One of the molecules of a triple-helical region often is an oligonucleotide.

As used herein, the term polynucleotide includes DNAs or RNAs as described above that contain one or more modified bases. Thus, DNAs or RNAs with backbones modified for stability or for other reasons are "polynucleotides" as that term is intended herein. Moreover, DNAs or RNAs comprising unusual bases, such as inosine, or modified bases, such as tritylated bases, to name just two examples, are polynucleotides as the term is used herein.

It will be appreciated that a great variety of modifications have been made to DNA and RNA that serve many useful purposes known to those of skill in the art. The term polynucleotide as it is employed herein embraces such chemically, enzymatically or metabolically modified forms of polynucleotides, as well as the chemical forms of DNA and RNA characteristic of viruses and cells, including simple and complex cells, inter alia.

POLYPEPTIDES, as used herein, includes all polypeptides as described below. The basic structure of polypeptides is well known and has been described in innumerable textbooks and other publications in the art. In this context, the term is used herein to refer to any peptide or protein comprising two or more amino acids joined to each other in a linear chain by peptide bonds. As used herein, the term refers to both short chains, which also commonly are referred to in the art as peptides, oligopeptides and oligomers, for example, and to longer chains, which generally are referred to in the art as proteins, of which there are many types.

It will be appreciated that polypeptides often contain amino acids other than the 20 amino acids commonly referred to as the 20 naturally occurring amino acids, and that many amino acids, including the terminal amino acids, may be modified in a given polypeptide, either by natural processes, such as processing and other post-translational modifications, but also by chemical modification techniques which are well known to the art. Even the common modifications that occur naturally in polypeptides are too numerous to list exhaustively here, but they are well described in basic texts and in more detailed monographs, as well as in a voluminous research literature, and they are well known to those of skill in the art. Among the known modifications which may be present in polypeptides of the present are, to name an illustrative few, acetylation, acylation, ADP-ribosylation, amidation, covalent attachment of flavin, covalent attachment of a heme moiety, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, covalent attachment of phosphatidylinositol, cross-linking, cyclization, disulfide bond formation, demethylation, formation of covalent cross-links, formation of cystine, formation of pyroglutamate, formylation, gamma-carboxylation, glycosylation, GPI anchor formation, hydroxylation, iodination, methylation, myristoylation, oxidation, proteolytic processing, phosphorylation, prenylation, racemization, selenoylation, sulfation, transfer-RNA mediated addition of amino acids to proteins such as arginylation, and ubiquitination.

Such modifications are well known to those of skill and have been described in great detail in the scientific literature. Several particularly common modifications,

glycosylation, lipid attachment, sulfation, gamma-carboxylation of glutamic acid residues, hydroxylation and ADP-ribosylation, for instance, are described in most basic texts, such as, for instance PROTEINS - STRUCTURE AND MOLECULAR PROPERTIES, 2nd Ed., T. E. Creighton, W. H. Freeman and Company, New York

5 (1993). Many detailed reviews are available on this subject, such as, for example, those provided by Wold, F., Posttranslational Protein Modifications: Perspectives and Prospects, pgs. 1-12 in POSTTRANSLATIONAL COVALENT MODIFICATION OF PROTEINS, B. C. Johnson, Ed., Academic Press, New York (1983); Seifter et al., Analysis for protein modifications and nonprotein cofactors,

10 Meth. Enzymol. 182: 626-646 (1990) and Rattan et al., Protein Synthesis: Posttranslational Modifications and Aging, Ann. N.Y. Acad. Sci. 663: 48-62 (1992).

It will be appreciated, as is well known and as noted above, that polypeptides are not always entirely linear. For instance, polypeptides may be branched as a result of ubiquitination, and they may be circular, with or without branching, generally as a

15 result of posttranslation events, including natural processing event and events brought about by human manipulation which do not occur naturally. Circular, branched and branched circular polypeptides may be synthesized by non-translation natural process and by entirely synthetic methods, as well.

Modifications can occur anywhere in a polypeptide, including the peptide

20 backbone, the amino acid side-chains and the amino or carboxyl termini. In fact, blockage of the amino or carboxyl group in a polypeptide, or both, by a covalent modification, is common in naturally occurring and synthetic polypeptides and such modifications may be present in polypeptides of the present invention, as well. For instance, the amino terminal residue of polypeptides made in *E. coli*, prior to

25 proteolytic processing, almost invariably will be N-formylmethionine.

The modifications that occur in a polypeptide often will be a function of how it is made. For polypeptides made by expressing a cloned gene in a host, for instance, the nature and extent of the modifications in large part will be determined by the host cell posttranslational modification capacity and the modification signals present in the

30 polypeptide amino acid sequence. For instance, as is well known, glycosylation often

does not occur in bacterial hosts such as *E. coli*. Accordingly, when glycosylation is desired, a polypeptide should be expressed in a glycosylating host, generally a eukaryotic cell. Insect cell often carry out the same posttranslational glycosylations as mammalian cells and, for this reason, insect cell expression systems have been developed to express efficiently mammalian proteins having native patterns of glycosylation, inter alia. Similar considerations apply to other modifications.

It will be appreciated that the same type of modification may be present in the same or varying degree at several sites in a given polypeptide. Also, a given polypeptide may contain many types of modifications.

In general, as used herein, the term polypeptide encompasses all such modifications, particularly those that are present in polypeptides synthesized by expressing a polynucleotide in a host cell.

VARIANT(S) of polynucleotides or polypeptides, as the term is used herein, are polynucleotides or polypeptides that differ from a reference polynucleotide or polypeptide, respectively. Variants in this sense are described below and elsewhere in the present disclosure in greater detail.

(1) A polynucleotide that differs in nucleotide sequence from another, reference polynucleotide. Generally, differences are limited so that the nucleotide sequences of the reference and the variant are closely similar overall and, in many regions, identical.

As noted below, changes in the nucleotide sequence of the variant may be silent. That is, they may not alter the amino acids encoded by the polynucleotide. Where alterations are limited to silent changes of this type a variant will encode a polypeptide with the same amino acid sequence as the reference. Also as noted below, changes in the nucleotide sequence of the variant may alter the amino acid sequence of a polypeptide encoded by the reference polynucleotide. Such nucleotide changes may result in amino acid substitutions, additions, deletions, fusions and truncations in the polypeptide encoded by the reference sequence, as discussed below.

(2) A polypeptide that differs in amino acid sequence from another, reference polypeptide. Generally, differences are limited so that the sequences of the reference and the variant are closely similar overall and, in many region, identical.

5 A variant and reference polypeptide may differ in amino acid sequence by one or more substitutions, additions, deletions, fusions and truncations, which may be present in any combination.

RECEPTOR MOLECULE, as used herein, refers to molecules which bind or interact specifically with human cytokine polypeptides of the present invention, including not only classic receptors, which are preferred, but also other molecules that specifically bind to or interact with polypeptides of the invention (which also may be referred to as "binding molecules" and "interaction molecules," respectively and as "human cytokine polypeptide binding molecules" and "human cytokine polypeptide interaction molecules." Binding between polypeptides of the invention and such molecules, including receptor or binding or interaction molecules may be exclusive to polypeptides of the invention, which is very highly preferred, or it may be highly specific for polypeptides of the invention, which is highly preferred, or it may be highly specific to a group of proteins that includes polypeptides of the invention, which is preferred, or it may be specific to several groups of proteins at least one of which includes polypeptides of the invention.

20 Receptors also may be non-naturally occurring, such as antibodies and antibody-derived reagents that bind specifically to polypeptides of the invention.

DESCRIPTION OF THE INVENTION

25 The present invention relates to novel human cytokine polypeptides and polynucleotides, among other things, as described in greater detail below. In particular, the invention relates to polypeptides and polynucleotides of a novel human cytokine polypeptide, which is related by amino acid sequence homology to TSG. The invention relates especially to human cytokine polypeptide having the nucleotide and amino acid sequences set out in Figure 1 (SEQ ID NO:2), and to the human cytokine polypeptide nucleotide and amino acid sequences of the human cDNA in the

30

deposited clone. It will be appreciated that the nucleotide and amino acid sequences set out in Figure 1 (SEQ ID NO:2) were obtained by sequencing the human cDNA of the deposited clone. Hence, the sequence of the deposited clone is controlling as to any discrepancies between the two and any reference to the sequence of Figure 1 (SEQ ID NO:1) includes reference to the sequence of the human cDNA of the deposited clone.

Polynucleotides

In accordance with one aspect of the present invention, there are provided isolated polynucleotides that encode the human cytokine polypeptide having the deduced amino acid sequence of Figure 1 (SEQ ID NO:2).

Using the information provided herein, such as the polynucleotide sequence set out in Figure 1 (SEQ ID NO:1), a polynucleotide of the present invention encoding a human cytokine polypeptide may be obtained using standard cloning and screening procedures, such as those for cloning cDNAs using mRNA from cells of human tissue as starting material. Illustrative of the invention, the polynucleotide set out in Figure 1 (SEQ ID NO:1) was discovered in a cDNA library derived from cells of human smooth muscle.

Human cytokine polypeptide of the invention is structurally related to other proteins of the connective tissue cytokine polypeptide family, as shown by the results of sequencing the human cDNA encoding human cytokine polypeptide in the deposited clone. The human cDNA sequence thus obtained is set out in Figure 1 (SEQ ID NO:1). It contains an open reading frame encoding a protein of about 223 amino acid residues having a putative 20 amino acid leader sequence. The full length protein has a deduced molecular weight of about 26 kDa. The protein exhibits greatest homology to Drosophila TSG protein among known proteins. The first 221 residues of the human cytokine polypeptide of Figure 1 (SEQ ID NO:2) have about 35.922% identity and about 60.680% similarity with the amino acid sequence of TSG.

Polynucleotides of the present invention may be in the form of RNA, such as mRNA, or in the form of DNA, including, for instance, cDNA and genomic DNA

obtained by cloning or produced by chemical synthetic techniques or by a combination thereof. The DNA may be double-stranded or single-stranded. Single-stranded DNA may be the coding strand, also known as the sense strand, or it may be the non-coding strand, also referred to as the anti-sense strand.

5 The coding sequence which encodes the polypeptide may be identical to the coding sequence of the polynucleotide shown in Figure 1 (SEQ ID NO:1). It also may be a polynucleotide with a different sequence, which, as a result of the redundancy (degeneracy) of the genetic code, encodes the polypeptide of the DNA of Figure 1 (SEQ ID NO:1).

10 Polynucleotides of the present invention which encode the polypeptide of Figure 1 (SEQ ID NO:2) may include, but are not limited to the coding sequence for the mature polypeptide, by itself; the coding sequence for the mature polypeptide and additional coding sequences, such as those encoding a leader or secretory sequence, such as a pre-, or pro- or prepro- protein sequence; the coding sequence of the mature
15 polypeptide, with or without the aforementioned additional coding sequences, together with additional, non-coding sequences, including for example, but not limited to introns and non-coding 5' and 3' sequences, such as the transcribed, non-translated sequences that play a role in transcription, mRNA processing - including splicing and polyadenylation signals, for example - ribosome binding and stability of mRNA;
20 additional coding sequence which codes for additional amino acids, such as those which provide additional functionalities. Thus, for instance, the polypeptide may be fused to a marker sequence, such as a peptide, which facilitates purification of the fused polypeptide. In certain preferred embodiments of this aspect of the invention, the marker sequence is a hexa-histidine peptide, such as the tag provided in a pQE
25 vector (Qiagen, Inc.), among others, many of which are commercially available. As described in Gentz et al., Proc. Natl. Acad. Sci., USA 86: 821-824 (1989), for instance, hexa-histidine provides for convenient purification of the fusion protein. The HA tag corresponds to an epitope derived of influenza hemagglutinin protein, which has been described by Wilson et al., Cell 37: 767 (1984), for instance.

In accordance with the foregoing, the term "polynucleotide encoding a polypeptide" as used herein encompasses polynucleotides which include a sequence encoding a polypeptide of the present invention, particularly the human cytokine polypeptide having the amino acid sequence set out in Figure 1 (SEQ ID NO:2). The term encompasses polynucleotides that include a single continuous region or discontinuous regions encoding the polypeptide (for example, interrupted by introns) together with additional regions.

The present invention further relates to variants of the herein above described polynucleotides which encode for fragments, analogs and derivatives of the polypeptide having the deduced amino acid sequence of Figure 1 (SEQ ID NO:2). A variant of the polynucleotide may be a naturally occurring variant such as a naturally occurring allelic variant, or it may be a variant that is not known to occur naturally. Such non-naturally occurring variants of the polynucleotide may be made by mutagenesis techniques, including those applied to polynucleotides, cells or organisms.

Among variants in this regard are variants that differ from the aforementioned polynucleotides by nucleotide substitutions, deletions or additions. The substitutions, deletions or additions may involve one or more nucleotides. The variants may be altered in coding or non-coding regions or both. Alterations in the coding regions may produce conservative or non-conservative amino acid substitutions, deletions or additions.

Among the particularly preferred embodiments of the invention in this regard are polynucleotides encoding polypeptides having the amino acid sequence of human cytokine polypeptide set out in Figure 1 (SEQ ID NO:2); variants, analogs, derivatives and fragments thereof, and fragments of the variants, analogs and derivatives.

Further particularly preferred in this regard are polynucleotides encoding human cytokine polypeptide variants, analogs, derivatives and fragments, and variants, analogs and derivatives of the fragments, which have the amino acid sequence of the human cytokine polypeptide of Figure 1 (SEQ ID NO:2) in which several, a

few, 5 to 10, 1 to 5, 1 to 3, 2, 1 or no amino acid residues are substituted, deleted or added, in any combination. Especially preferred among these are silent substitutions, additions and deletions, which do not alter the properties and activities of the human cytokine polypeptide. Also especially preferred in this regard are conservative substitutions. Most highly preferred are polynucleotides encoding polypeptides having the amino acid sequence of Figure 1 (SEQ ID NO:2) without substitutions.

Further preferred embodiments of the invention are polynucleotides that are at least 70% identical to a polynucleotide encoding the human cytokine polypeptide having the amino acid sequence set out in Figure 1 (SEQ ID NO:2), and polynucleotides which are complementary to such polynucleotides. Alternatively, most highly preferred are polynucleotides that comprise a region that is at least 80% identical to a polynucleotide encoding the human cytokine polypeptide and polynucleotides complementary thereto. In this regard, polynucleotides at least 90% identical to the same are particularly preferred, and among these particularly preferred polynucleotides, those with at least 95% are especially preferred. Furthermore, those with at least 97% are highly preferred among those with at least 95%, and among these those with at least 98% and at least 99% are particularly highly preferred, with at least 99% being the more preferred.

Particularly preferred embodiments in this respect, moreover, are polynucleotides which encode polypeptides which retain substantially the same biological function or activity as the mature polypeptide encoded by the human cDNA of Figure 1 (SEQ ID NO:1).

The present invention further relates to polynucleotides that hybridize to the herein above-described sequences. In this regard, the present invention especially relates to polynucleotides which hybridize under stringent conditions to the herein above-described polynucleotides. As herein used, the term "stringent conditions" means hybridization will occur only if there is at least 95% and preferably at least 97% identity between the sequences.

As discussed additionally herein regarding polynucleotide assays of the invention, for instance, polynucleotides of the invention as discussed above, may be

used as a hybridization probe for cDNA and genomic DNA to isolate full-length cDNAs and genomic clones encoding human cytokine polypeptide and to isolate cDNA and genomic clones of other genes that have a high sequence similarity to the human cytokine polypeptide gene. Such probes generally will comprise at least 15
5 bases. Preferably, such probes will have at least 30 bases and may have at least 50 bases. Particularly preferred probes will have at least 30 bases and will have 50 bases or less.

For example, the coding region of the human cytokine polypeptide gene may be isolated by screening using the known DNA sequence to synthesize an
10 oligonucleotide probe. A labeled oligonucleotide having a sequence complementary to that of a gene of the present invention is then used to screen a library of human cDNA, genomic DNA or mRNA to determine which members of the library the probe hybridizes to.

The polynucleotides and polypeptides of the present invention may be
15 employed as research reagents and materials for discovery of treatments and diagnostics to human disease, as further discussed herein relating to polynucleotide assays, inter alia.

The polynucleotides may encode a polypeptide which is the mature protein plus additional amino or carboxyl-terminal amino acids, or amino acids interior to the
20 mature polypeptide (when the mature form has more than one polypeptide chain, for instance). Such sequences may play a role in processing of a protein from precursor to a mature form, may facilitate protein trafficking, may prolong or shorten protein half-life or may facilitate manipulation of a protein for assay or production, among other things. As generally is the case in situ, the additional amino acids may be
25 processed away from the mature protein by cellular enzymes.

A precursor protein, having the mature form of the polypeptide fused to one or more prosequences may be an inactive form of the polypeptide. When prosequences are removed such inactive precursors generally are activated. Some or all of the prosequences may be removed before activation. Generally, such precursors
30 are called proproteins.

In sum, a polynucleotide of the present invention may encode a mature protein, a mature protein plus a leader sequence (which may be referred to as a preprotein), a precursor of a mature protein having one or more prosequences which are not the leader sequences of a preprotein, or a preproprotein, which is a precursor to a proprotein, having a leader sequence and one or more prosequences, which generally are removed during processing steps that produce active and mature forms of the polypeptide.

Deposited materials

10 A deposit containing a human cytokine polypeptide cDNA has been deposited with the American Type Culture Collection, as noted above. Also as noted above, the cDNA deposit is referred to herein as "the deposited clone" or as "the cDNA of the deposited clone."

15 The deposited clone was deposited with the American Type Culture Collection, 12301 Park Lawn Drive, Rockville, Maryland 20852, USA, on March 19, 1996 and assigned ATCC Deposit No. 97486.

The deposited material is a baculovirus expression vector, pA2, that contains the full length human cytokine polypeptide cDNA, referred to as "PF270" upon deposit.

20 The deposit has been made under the terms of the Budapest Treaty on the international recognition of the deposit of micro-organisms for purposes of patent procedure. The strain will be irrevocably and without restriction or condition released to the public upon the issuance of a patent. The deposit is provided merely as convenience to those of skill in the art and is not an admission that a deposit is required for enablement, such as that required under 35 U.S.C. 112.

25 The sequence of the polynucleotides contained in the deposited material, as well as the amino acid sequence of the polypeptide encoded thereby, are controlling in the event of any conflict with any description of sequences herein.

A license may be required to make, use or sell the deposited materials, and no such license is hereby granted.

Polypeptides

The present invention further relates to a human cytokine polypeptide which has the deduced amino acid sequence of Figure 1 (SEQ ID NO:2).

5 The invention also relates to fragments, analogs and derivatives of these polypeptides. The terms "fragment," "derivative" and "analog" when referring to the polypeptide of Figure 1 (SEQ ID NO:2) means a polypeptide which retains essentially the same biological function or activity as such polypeptide. Thus, an analog includes a proprotein which can be activated by cleavage of the proprotein
10 portion to produce an active mature polypeptide.

The polypeptide of the present invention may be a recombinant polypeptide, a natural polypeptide or a synthetic polypeptide. In certain preferred embodiments it is a recombinant polypeptide.

15 The fragment, derivative or analog of the polypeptide of Figure 1 (SEQ ID NO:2) may be (i) one in which one or more of the amino acid residues are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code, or (ii) one in which one or more of the amino acid residues includes a substituent group, or (iii) one in which the mature polypeptide is fused
20 with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol), or (iv) one in which the additional amino acids are fused to the mature polypeptide, such as a leader or secretory sequence or a sequence which is employed for purification of the mature polypeptide or a proprotein sequence. Such fragments, derivatives and analogs are deemed to be
25 within the scope of those skilled in the art from the teachings herein.

Among the particularly preferred embodiments of the invention in this regard are polypeptides having the amino acid sequence of human cytokine polypeptide set out in Figure 1 (SEQ ID NO:2), variants, analogs, derivatives and fragments thereof, and variants, analogs and derivatives of the fragments. Alternatively, particularly
30 preferred embodiments of the invention in this regard are polypeptides having the

amino acid sequence of the human cytokine polypeptide of the cDNA in the deposited clone, variants, analogs, derivatives and fragments thereof, and variants, analogs and derivatives of the fragments.

Among preferred variants are those that vary from a reference by conservative amino acid substitutions. Such substitutions are those that substitute a given amino acid in a polypeptide by another amino acid of like characteristics. Typically seen as conservative substitutions are the replacements, one for another, among the aliphatic amino acids Ala, Val, Leu and Ile; interchange of the hydroxyl residues Ser and Thr, exchange of the acidic residues Asp and Glu, substitution between the amide residues Asn and Gln, exchange of the basic residues Lys and Arg and replacements among the aromatic residues Phe, Tyr.

Further particularly preferred in this regard are variants, analogs, derivatives and fragments, and variants, analogs and derivatives of the fragments, having the amino acid sequence of the human cytokine polypeptide of Figure 1 (SEQ ID NO:2) in which several, a few, 5 to 10, 1 to 5, 1 to 3, 2, 1 or no amino acid residues are substituted, deleted or added, in any combination. Especially preferred among these are silent substitutions, additions and deletions, which do not alter the properties and activities of the human cytokine polypeptide. Also especially preferred in this regard are conservative substitutions. Most highly preferred are polypeptides having the amino acid sequence of Figure 1 (SEQ ID NO:2) without substitutions.

The polypeptides and polynucleotides of the present invention are preferably provided in an isolated form, and preferably are purified to homogeneity.

The polypeptides of the present invention also include the polypeptide of SEQ ID NO:2 (in particular the mature polypeptide) as well as polypeptides which have at least 70% similarity (preferably at least 70% identity) to the polypeptide of SEQ ID NO:2 and more preferably at least 90% similarity (more preferably at least 90% identity) to the polypeptide of SEQ ID NO:2 and still more preferably at least 95% similarity (still more preferably at least 95% identity) to the polypeptide of SEQ ID NO:2 and also include portions of such polypeptides with such portion of the

polypeptide generally containing at least 30 amino acids and more preferably at least 50 amino acids.

As known in the art "similarity" between two polypeptides is determined by comparing the amino acid sequence and its conserved amino acid substitutes of one
5 polypeptide to the sequence of a second polypeptide.

Fragments or portions of the polypeptides of the present invention may be employed for producing the corresponding full-length polypeptide by peptide synthesis; therefore, the fragments may be employed as intermediates for producing the full-length polypeptides. Fragments or portions of the polynucleotides of the
10 present invention may be used to synthesize full-length polynucleotides of the present invention.

Fragments

Also among preferred embodiments of this aspect of the present invention are
15 polypeptides comprising fragments of human cytokine polypeptide, most particularly fragments of the human cytokine polypeptide having the amino acid set out in Figure 1 (SEQ ID NO:2), and fragments of variants and derivatives of the human cytokine polypeptide of Figure 1 (SEQ ID NO:2).

In this regard a fragment is a polypeptide having an amino acid sequence that
20 entirely is the same as part but not all of the amino acid sequence of the aforementioned human cytokine polypeptide and variants or derivatives thereof.

Such fragments may be "free-standing," i.e., not part of or fused to other amino acids or polypeptides, or they may be comprised within a larger polypeptide of which they form a part or region. When comprised within a larger polypeptide, the
25 presently discussed fragments most preferably form a single continuous region. However, several fragments may be comprised within a single larger polypeptide. For instance, certain preferred embodiments relate to a fragment of a human cytokine polypeptide of the present comprised within a precursor polypeptide designed for expression in a host and having heterologous pre and pro-polypeptide regions fused to
30 the amino terminus of the human cytokine polypeptide fragment and an additional

region fused to the carboxyl terminus of the fragment. Therefore, fragments in one aspect of the meaning intended herein, refers to the portion or portions of a fusion polypeptide or fusion protein derived from human cytokine polypeptide.

As representative examples of polypeptide fragments of the invention, there
5 may be mentioned those which have from about 150 to about 223 amino acids.

In this context about includes the particularly recited range and ranges larger or smaller by several, a few, 5, 4, 3, 2 or 1 amino acid at either extreme or at both extremes. For instance, about 150 amino acids in this context means a polypeptide fragment of 150 amino acids plus or minus several, a few, 5, 4, 3, 2 or 1 amino acids to
10 15 plus or minus several a few, 5, 4, 3, 2 or 1 amino acid residues, i.e., ranges as broad as 150 minus several amino acids to 223 plus several amino acids to as narrow as 150 plus several amino acids to 170 minus several amino acids.

Highly preferred in this regard are the recited ranges plus or minus as many as 5 amino acids at either or at both extremes. Particularly highly preferred are the
15 recited ranges plus or minus as many as 3 amino acids at either or at both the recited extremes. Especially particularly highly preferred are ranges plus or minus 1 amino acid at either or at both extremes or the recited ranges with no additions or deletions. Most highly preferred of all in this regard are fragments from about 203 to about 223 amino acids.

Among especially preferred fragments of the invention are truncation mutants
20 of human cytokine polypeptide. Truncation mutants include human cytokine polypeptides having the amino acid sequence of Figure 1 (SEQ ID NO:2), or of variants or derivatives thereof, except for deletion of a continuous series of residues (that is, a continuous region, part or portion) that includes the amino terminus, or a
25 continuous series of residues that includes the carboxyl terminus or, as in double truncation mutants, deletion of two continuous series of residues, one including the amino terminus and one including the carboxyl terminus. Fragments having the size ranges set out about also are preferred embodiments of truncation fragments, which are especially preferred among fragments generally.

Also preferred in this aspect of the invention are fragments characterized by structural or functional attributes of human cytokine polypeptide. Preferred embodiments of the invention in this regard include fragments that comprise alpha-helix and alpha-helix forming regions ("alpha-regions"), beta-sheet and beta-sheet-forming regions ("beta-regions"), turn and turn-forming regions ("turn-regions"), coil and coil-forming regions ("coil-regions"), hydrophilic regions, hydrophobic regions, alpha amphipathic regions, beta amphipathic regions, flexible regions, surface-forming regions and high antigenic index regions of human cytokine polypeptide.

Certain preferred regions in these regards include, but are not limited to, regions of the aforementioned types identified by analysis of the amino acid sequence set out in Figure 1 (SEQ ID NO:2). Such preferred regions include Garnier-Robson alpha-regions, beta-regions, turn-regions and coil-regions, Chou-Fasman alpha-regions, beta-regions and turn-regions, Kyte-Doolittle hydrophilic regions and hydrophilic regions, Eisenberg alpha and beta amphipathic regions, Karplus-Schulz flexible regions, Emini surface-forming regions and Jameson-Wolf high antigenic index regions.

Among highly preferred fragments in this regard are those that comprise regions of human cytokine polypeptide that combine several structural features, such as several of the features set out above. In this regard, the regions defined by the residues about 203 to about 226 of Figure 1 (SEQ ID NO:2), which all are characterized by amino acid compositions highly characteristic of turn-regions, hydrophilic regions, flexible-regions, surface-forming regions, and high antigenic index-regions, are especially highly preferred regions. Such regions may be comprised within a larger polypeptide or may be by themselves a preferred fragment of the present invention, as discussed above. It will be appreciated that the term "about" as used in this paragraph has the meaning set out above regarding fragments in general.

Further preferred regions are those that mediate activities of human cytokine polypeptide. Most highly preferred in this regard are fragments that have a chemical, biological or other activity of human cytokine polypeptide, including those with a similar activity or an improved activity, or with a decreased undesirable activity. Highly preferred in this regard are fragments that contain regions that are homologs in

sequence, or in position, or in both sequence and to active regions of related polypeptides, such as the related polypeptides set out in Figure 2 (SEQ ID NO:9), which include *tsg*. Among particularly preferred fragments in these regards are truncation mutants, as discussed above.

5 It will be appreciated that the invention also relates to, among others, polynucleotides encoding the aforementioned fragments, polynucleotides that hybridize to polynucleotides encoding the fragments, particularly those that hybridize under stringent conditions, and polynucleotides, such as PCR primers, for amplifying polynucleotides that encode the fragments. In these regards, preferred
10 polynucleotides are those that correspondent to the preferred fragments, as discussed above.

Vectors, host cells, expression

The present invention also relates to vectors which include polynucleotides of the present invention, host cells which are genetically engineered with vectors of the
15 invention and the production of polypeptides of the invention by recombinant techniques.

Host cells can be genetically engineered to incorporate polynucleotides and express polypeptides of the present invention. For instance, polynucleotides may be introduced into host cells using well known techniques of infection, transduction,
20 transfection, transvection and transformation. The polynucleotides may be introduced alone or with other polynucleotides. Such other polynucleotides may be introduced independently, co-introduced or introduced joined to the polynucleotides of the invention.

Thus, for instance, polynucleotides of the invention may be transfected into
25 host cells with another, separate, polynucleotide encoding a selectable marker, using standard techniques for co-transfection and selection in, for instance, mammalian cells. In this case the polynucleotides generally will be stably incorporated into the host cell genome.

Alternatively, the polynucleotides may be joined to a vector containing a
30 selectable marker for propagation in a host. The vector construct may be introduced

into host cells by the aforementioned techniques. Generally, a plasmid vector is introduced as DNA in a precipitate, such as a calcium phosphate precipitate, or in a complex with a charged lipid. Electroporation also may be used to introduce polynucleotides into a host. If the vector is a virus, it may be packaged in vitro or introduced into a packaging cell and the packaged virus may be transduced into cells. A wide variety of techniques suitable for making polynucleotides and for introducing polynucleotides into cells in accordance with this aspect of the invention are well known and routine to those of skill in the art. Such techniques are reviewed at length in Sambrook et al. cited above, which is illustrative of the many laboratory manuals that detail these techniques. In accordance with this aspect of the invention the vector may be, for example, a plasmid vector, a single or double-stranded phage vector, a single or double-stranded RNA or DNA viral vector. Such vectors may be introduced into cells as polynucleotides, preferably DNA, by well known techniques for introducing DNA and RNA into cells. The vectors, in the case of phage and viral vectors also may be and preferably are introduced into cells as packaged or encapsidated virus by well known techniques for infection and transduction. Viral vectors may be replication competent or replication defective. In the latter case viral propagation generally will occur only in complementing host cells.

Preferred among vectors, in certain respects, are those for expression of polynucleotides and polypeptides of the present invention. Generally, such vectors comprise cis-acting control regions effective for expression in a host operatively linked to the polynucleotide to be expressed. Appropriate trans-acting factors either are supplied by the host, supplied by a complementing vector or supplied by the vector itself upon introduction into the host.

In certain preferred embodiments in this regard, the vectors provide for specific expression. Such specific expression may be inducible expression or expression only in certain types of cells or both inducible and cell-specific. Particularly preferred among inducible vectors are vectors that can be induced for expression by environmental factors that are easy to manipulate, such as temperature and nutrient additives. A variety of vectors suitable to this aspect of the invention,

including constitutive and inducible expression vectors for use in prokaryotic and eukaryotic hosts, are well known and employed routinely by those of skill in the art.

5 The engineered host cells can be cultured in conventional nutrient media, which may be modified as appropriate for, inter alia, activating promoters, selecting transformants or amplifying genes. Culture conditions, such as temperature, pH and the like, previously used with the host cell selected for expression generally will be suitable for expression of polypeptides of the present invention as will be apparent to those of skill in the art.

10 A great variety of expression vectors can be used to express a polypeptide of the invention. Such vectors include chromosomal, episomal and virus-derived vectors e.g., vectors derived from bacterial plasmids, from bacteriophage, from yeast episomes, from yeast chromosomal elements, from viruses such as baculoviruses, papova viruses, such as SV40, vaccinia viruses, adenoviruses, fowl pox viruses, pseudorabies viruses and retroviruses, and vectors derived from combinations thereof, 15 such as those derived from plasmid and bacteriophage genetic elements, such as cosmids and phagemids, all may be used for expression in accordance with this aspect of the present invention. Generally, any vector suitable to maintain, propagate or express polynucleotides to express a polypeptide in a host may be used for expression in this regard.

20 The appropriate DNA sequence may be inserted into the vector by any of a variety of well-known and routine techniques. In general, a DNA sequence for expression is joined to an expression vector by cleaving the DNA sequence and the expression vector with one or more restriction endonucleases and then joining the restriction fragments together using T4 DNA ligase. Procedures for restriction and 25 ligation that can be used to this end are well known and routine to those of skill. Suitable procedures in this regard, and for constructing expression vectors using alternative techniques, which also are well known and routine to those skill, are set forth in great detail in Sambrook et al. cited elsewhere herein.

30 The DNA sequence in the expression vector is operatively linked to appropriate expression control sequence(s), including, for instance, a promoter to

direct mRNA transcription. Representatives of such promoters include the phage lambda PL promoter, the E. coli lac, trp and tac promoters, the SV40 early and late promoters and promoters of retroviral LTRs, to name just a few of the well-known promoters. It will be understood that numerous promoters not mentioned are suitable for use in this aspect of the invention are well known and readily may be employed by those of skill in the manner illustrated by the discussion and the examples herein.

In general, expression constructs will contain sites for transcription initiation and termination, and, in the transcribed region, a ribosome binding site for translation. The coding portion of the mature transcripts expressed by the constructs will include a translation initiating AUG at the beginning and a termination codon appropriately positioned at the end of the polypeptide to be translated.

In addition, the constructs may contain control regions that regulate as well as engender expression. Generally, in accordance with many commonly practiced procedures, such regions will operate by controlling transcription, such as repressor binding sites and enhancers, among others.

Vectors for propagation and expression generally will include selectable markers. Such markers also may be suitable for amplification or the vectors may contain additional markers for this purpose. In this regard, the expression vectors preferably contain one or more selectable marker genes to provide a phenotypic trait for selection of transformed host cells. Preferred markers include dihydrofolate reductase or neomycin resistance for eukaryotic cell culture, and tetracycline, theomycin, kanamycin or ampicillin resistance genes for culturing E. coli and other bacteria.

The vector containing the appropriate DNA sequence as described elsewhere herein, as well as an appropriate promoter, and other appropriate control sequences, may be introduced into an appropriate host using a variety of well known techniques suitable to expression therein of a desired polypeptide. Representative examples of appropriate hosts include bacterial cells, such as E. coli, Streptomyces and Salmonella typhimurium cells; fungal cells, such as yeast cells; insect cells such as Drosophila S2 and Spodoptera Sf9 cells; animal cells such as CHO, COS and Bowes melanoma cells;

and plant cells. Hosts for of a great variety of expression constructs are well known, and those of skill will be enabled by the present disclosure readily to select a host for expressing a polypeptides in accordance with this aspect of the present invention.

Various mammalian cell culture systems can be employed for expression, as
5 well. Examples of mammalian expression systems include the COS-7 lines of monkey kidney fibroblast, described in Gluzman et al., Cell 23: 175 (1981). Other cell lines capable of expressing a compatible vector include for example, the C127, 3T3, CHO, HeLa, human kidney 293 and BHK cell lines.

More particularly, the present invention also includes recombinant constructs,
10 such as expression constructs, comprising one or more of the sequences described above. The constructs comprise a vector, such as a plasmid or viral vector, into which such a sequence of the invention has been inserted. The sequence may be inserted in a forward or reverse orientation. In certain preferred embodiments in this regard, the construct further comprises regulatory sequences, including, for example, a promoter,
15 operably linked to the sequence. Large numbers of suitable vectors and promoters are known to those of skill in the art, and there are many commercially available vectors suitable for use in the present invention.

The following vectors, which are commercially available, are provided by way of example. Among vectors preferred for use in bacteria are pQE70, pQE60 and pQE-
20 9, available from Qiagen; pBS vectors, Phagescript vectors, Bluescript vectors, pNH8A, pNH16a, pNH18A, pNH46A, available from Stratagene; and ptrc99a, pKK223-3, pKK223-3, pDR540, pRIT5 available from Pharmacia. Among preferred eukaryotic vectors are pWLNEO, pSV2CAT, pOG44, pXT1 and pSG available from Stratagene; and pSVK3, pBPV, pMSG and pSVL available from Pharmacia. These
25 vectors are listed solely by way of illustration of the many commercially available and well known vectors that are available to those of skill in the art for use in accordance with this aspect of the present invention. It will be appreciated that any other plasmid or vector suitable for, for example, introduction, maintenance, propagation or expression of a polynucleotide or polypeptide of the invention in a host may be used
30 in this aspect of the invention.

Promoter regions can be selected from any desired gene using vectors that contain a reporter transcription unit lacking a promoter region, such as a chloramphenicol acetyl transferase ("cat") transcription unit, downstream of restriction site or sites for introducing a candidate promoter fragment; i.e., a fragment
5 that may contain a promoter. As is well known, introduction into the vector of a promoter-containing fragment at the restriction site upstream of the cat gene engenders production of CAT activity, which can be detected by standard CAT assays. Vectors suitable to this end are well known and readily available. Two such vectors are pKK232-8 and pCM7. Thus, promoters for expression of polynucleotides of the
10 present invention include not only well known and readily available promoters, but also promoters that readily may be obtained by the foregoing technique, using a reporter gene.

Among known bacterial promoters suitable for expression of polynucleotides and polypeptides in accordance with the present invention are the E. coli lacI and lacZ
15 promoters, the T3 and T7 promoters, the T5 tac promoter, the lambda PR, PL promoters and the trp promoter. Among known eukaryotic promoters suitable in this regard are the CMV immediate early promoter, the HSV thymidine kinase promoter, the early and late SV40 promoters, the promoters of retroviral LTRs, such as those of the Rous sarcoma virus ("RSV"), and metallothionein promoters, such as the mouse
20 metallothionein-I promoter.

Selection of appropriate vectors and promoters for expression in a host cell is a well known procedure and the requisite techniques for expression vector construction, introduction of the vector into the host and expression in the host are routine skills in the art.

25 Generally, recombinant expression vectors will include origins of replication, a promoter derived from a highly-expressed gene to direct transcription of a downstream structural sequence, and a selectable marker to permit isolation of vector containing cells after exposure to the vector.

The present invention also relates to host cells containing the above-described
30 constructs discussed above. The host cell can be a higher eukaryotic cell, such as a

mammalian cell, or a lower eukaryotic cell, such as a yeast cell, or the host cell can be a prokaryotic cell, such as a bacterial cell.

Constructs in host cells can be used in a conventional manner to produce the gene product encoded by the recombinant sequence. Alternatively, the polypeptides
5 of the invention can be synthetically produced by conventional peptide synthesizers.

Mature proteins can be expressed in mammalian cells, yeast, bacteria, or other cells under the control of appropriate promoters. Cell-free translation systems can also be employed to produce such proteins using RNAs derived from the DNA constructs of the present invention. Appropriate cloning and expression vectors for
10 use with prokaryotic and eukaryotic hosts are described by Sambrook et al., MOLECULAR CLONING: A LABORATORY MANUAL, 2nd Ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989).

Transcription of the DNA encoding the polypeptides of the present invention by higher eukaryotes may be increased by inserting an enhancer sequence into the
15 vector. Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp that act to increase transcriptional activity of a promoter in a given host cell-type. Examples of enhancers include the SV40 enhancer, which is located on the late side of the replication origin at bp 100 to 270, the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus
20 enhancers.

Polynucleotides of the invention, encoding the heterologous structural sequence of a polypeptide of the invention generally will be inserted into the vector using standard techniques so that it is operably linked to the promoter for expression. The polynucleotide will be positioned so that the transcription start site is located
25 appropriately 5' to a ribosome binding site. The ribosome binding site will be 5' to the AUG that initiates translation of the polypeptide to be expressed. Generally, there will be no other open reading frames that begin with an initiation codon, usually AUG, and lie between the ribosome binding site and the initiating AUG. Also, generally, there will be a translation stop codon at the end of the polypeptide and

there will be a polyadenylation signal and a transcription termination signal appropriately disposed at the 3' end of the transcribed region.

For secretion of the translated protein into the lumen of the endoplasmic reticulum, into the periplasmic space or into the extracellular environment, appropriate secretion signals may be incorporated into the expressed polypeptide. The signals may be endogenous to the polypeptide or they may be heterologous signals.

The polypeptide may be expressed in a modified form, such as a fusion protein, and may include not only secretion signals but also additional heterologous functional regions. Thus, for instance, a region of additional amino acids, particularly charged amino acids, may be added to the N-terminus of the polypeptide to improve stability and persistence in the host cell, during purification or during subsequent handling and storage. Also, region also may be added to the polypeptide to facilitate purification. Such regions may be removed prior to final preparation of the polypeptide. The addition of peptide moieties to polypeptides to engender secretion or excretion, to improve stability and to facilitate purification, among others, are familiar and routine techniques in the art.

Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, where the selected promoter is inducible it is induced by appropriate means (e.g., temperature shift or exposure to chemical inducer) and cells are cultured for an additional period.

Cells typically then are harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract retained for further purification.

Microbial cells employed in expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents, such methods are well know to those skilled in the art.

The human cytokine polypeptide can be recovered and purified from recombinant cell cultures by well-known methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity

chromatography, hydroxylapatite chromatography and lectin chromatography. Most preferably, high performance liquid chromatography ("HPLC") is employed for purification. Well known techniques for refolding protein may be employed to regenerate active conformation when the polypeptide is denatured during isolation and or purification.

Polypeptides of the present invention include naturally purified products, products of chemical synthetic procedures, and products produced by recombinant techniques from a prokaryotic or eukaryotic host, including, for example, bacterial, yeast, higher plant, insect and mammalian cells. Depending upon the host employed in a recombinant production procedure, the polypeptides of the present invention may be glycosylated or may be non-glycosylated. In addition, polypeptides of the invention may also include an initial modified methionine residue, in some cases as a result of host-mediated processes.

human cytokine polypeptide polynucleotides and polypeptides may be used in accordance with the present invention for a variety of applications, particularly those that make use of the chemical and biological properties of human cytokine polypeptide. Among these are applications in cell proliferation and differentiation. Additional applications relate to diagnosis and to treatment of disorders of cells, tissues and organisms. These aspects of the invention are illustrated further by the following discussion.

Polynucleotide assays

This invention is also related to the use of the human cytokine polypeptide polynucleotides to detect complementary polynucleotides such as, for example, as a diagnostic reagent. Detection of a mutated form of human cytokine polypeptide associated with a dysfunction will provide a diagnostic tool that can add or define a diagnosis of a disease or susceptibility to a disease which results from under-expression over-expression or altered expression of human cytokine polypeptide, such as, for example, neoplasia.

Individuals carrying mutations in the human cytokine polypeptide gene may be detected at the DNA level by a variety of techniques. Nucleic acids for diagnosis may be obtained from a patient's cells, such as from blood, urine, saliva, tissue biopsy and autopsy material. The genomic DNA may be used directly for detection or may
5 be amplified enzymatically by using PCR prior to analysis. PCR (Saiki et al., Nature, 324: 163-166 (1986)). RNA or cDNA may also be used in the same ways. As an example, PCR primers complementary to the nucleic acid encoding human cytokine polypeptide can be used to identify and analyze human cytokine polypeptide expression and mutations. For example, deletions and insertions can be detected by a
10 change in size of the amplified product in comparison to the normal genotype. Point mutations can be identified by hybridizing amplified DNA to radiolabeled human cytokine polypeptide RNA or alternatively, radiolabeled human cytokine polypeptide antisense DNA sequences. Perfectly matched sequences can be distinguished from mismatched duplexes by RNase A digestion or by differences in
15 melting temperatures.

Sequence differences between a reference gene and genes having mutations also may be revealed by direct DNA sequencing. In addition, cloned DNA segments may be employed as probes to detect specific DNA segments. The sensitivity of such methods can be greatly enhanced by appropriate use of PCR or another amplification
20 method. For example, a sequencing primer is used with double-stranded PCR product or a single-stranded template molecule generated by a modified PCR. The sequence determination is performed by conventional procedures with radiolabeled nucleotide or by automatic sequencing procedures with fluorescent-tags.

Genetic testing based on DNA sequence differences may be achieved by
25 detection of alteration in electrophoretic mobility of DNA fragments in gels, with or without denaturing agents. Small sequence deletions and insertions can be visualized by high resolution gel electrophoresis. DNA fragments of different sequences may be distinguished on denaturing formamide gradient gels in which the mobilities of different DNA fragments are retarded in the gel at different positions according to

their specific melting or partial melting temperatures (see, e.g., Myers et al., Science, 230: 1242 (1985)).

Sequence changes at specific locations also may be revealed by nuclease protection assays, such as RNase and S1 protection or the chemical cleavage method
5 (e.g., Cotton et al., Proc. Natl. Acad. Sci., USA, 85: 4397-4401 (1985)).

Thus, the detection of a specific DNA sequence may be achieved by methods such as hybridization, RNase protection, chemical cleavage, direct DNA sequencing or the use of restriction enzymes, (e.g., restriction fragment length polymorphisms ("RFLP") and Southern blotting of genomic DNA.

10 In addition to more conventional gel-electrophoresis and DNA sequencing, mutations also can be detected by in situ analysis.

Chromosome assays

The sequences of the present invention are also valuable for chromosome
15 identification. The sequence is specifically targeted to and can hybridize with a particular location on an individual human chromosome. Moreover, there is a current need for identifying particular sites on the chromosome. Few chromosome marking reagents based on actual sequence data (repeat polymorphisms) are presently available for marking chromosomal location. The mapping of DNAs to chromosomes according
20 to the present invention is an important first step in correlating those sequences with genes associated with disease.

In certain preferred embodiments in this regard, the cDNA herein disclosed is used to clone genomic DNA of a human cytokine polypeptide gene. This can be accomplished using a variety of well known techniques and libraries, which generally
25 are available commercially. The genomic DNA the is used for in situ chromosome mapping using well known techniques for this purpose. Typically, in accordance with routine procedures for chromosome mapping, some trial and error may be necessary to identify a genomic probe that gives a good in situ hybridization signal.

In some cases, in addition, sequences can be mapped to chromosomes by
30 preparing PCR primers (preferably 15-25 bp) from the cDNA. Computer analysis of

the 3' untranslated region of the gene is used to rapidly select primers that do not span more than one exon in the genomic DNA, thus complicating the amplification process. These primers are then used for PCR screening of somatic cell hybrids containing individual human chromosomes. Only those hybrids containing the human gene
5 corresponding to the primer will yield an amplified fragment.

PCR mapping of somatic cell hybrids is a rapid procedure for assigning a particular DNA to a particular chromosome. Using the present invention with the same oligonucleotide primers, sublocalization can be achieved with panels of fragments from specific chromosomes or pools of large genomic clones in an analogous
10 manner. Other mapping strategies that can similarly be used to map to its chromosome include in situ hybridization, prescreening with labeled flow-sorted chromosomes and preselection by hybridization to construct chromosome specific-cDNA libraries.

Fluorescence in situ hybridization ("FISH") of a cDNA clone to a metaphase
15 chromosomal spread can be used to provide a precise chromosomal location in one step. This technique can be used with cDNA as short as 50 or 60 bases. For a review of this technique, see Verma et al., HUMAN CHROMOSOMES: A MANUAL OF BASIC TECHNIQUES, Pergamon Press, New York (1988).

Once a sequence has been mapped to a precise chromosomal location, the
20 physical position of the sequence on the chromosome can be correlated with genetic map data. Such data are found, for example, in V. McKusick, MENDELIAN INHERITANCE IN MAN, available on line through Johns Hopkins University, Welch Medical Library. The relationship between genes and diseases that have been mapped to the same chromosomal region are then identified through linkage analysis
25 (coinheritance of physically adjacent genes).

Next, it is necessary to determine the differences in the cDNA or genomic sequence between affected and unaffected individuals. If a mutation is observed in some or all of the affected individuals but not in any normal individuals, then the mutation is likely to be the causative agent of the disease.

With current resolution of physical mapping and genetic mapping techniques, a cDNA precisely localized to a chromosomal region associated with the disease could be one of between 50 and 500 potential causative genes. (This assumes 1 megabase mapping resolution and one gene per 20 kb).

5

Polypeptide assays

The present invention also relates to a diagnostic assays such as quantitative and diagnostic assays for detecting levels of human cytokine polypeptide protein in cells and tissues, including determination of normal and abnormal levels. Thus, for instance, a diagnostic assay in accordance with the invention for detecting over-expression of human cytokine polypeptide protein compared to normal control tissue samples may be used to detect the presence of neoplasia, for example. Assay techniques that can be used to determine levels of a protein, such as an human cytokine polypeptide protein of the present invention, in a sample derived from a host are well-known to those of skill in the art. Such assay methods include radioimmunoassays, competitive-binding assays, Western Blot analysis and ELISA assays. Among these ELISAs frequently are preferred. An ELISA assay initially comprises preparing an antibody specific to human cytokine polypeptide, preferably a monoclonal antibody. In addition a reporter antibody generally is prepared which binds to the monoclonal antibody. The reporter antibody is attached a detectable reagent such as radioactive, fluorescent or enzymatic reagent, in this example horseradish peroxidase enzyme.

To carry out an ELISA a sample is removed from a host and incubated on a solid support, e.g. a polystyrene dish, that binds the proteins in the sample. Any free protein binding sites on the dish are then covered by incubating with a non-specific protein such as bovine serum albumin. Next, the monoclonal antibody is incubated in the dish during which time the monoclonal antibodies attach to any human cytokine polypeptide proteins attached to the polystyrene dish. Unbound monoclonal antibody is washed out with buffer. The reporter antibody linked to horseradish peroxidase is placed in the dish resulting in binding of the reporter antibody to any

monoclonal antibody bound to human cytokine polypeptide. Unattached reporter antibody is then washed out. Reagents for peroxidase activity, including a colorimetric substrate are then added to the dish. Immobilized peroxidase, linked to human cytokine polypeptide through the primary and secondary antibodies, produces a colored reaction product. The amount of color developed in a given time period indicates the amount of human cytokine polypeptide protein present in the sample. Quantitative results typically are obtained by reference to a standard curve.

A competition assay may be employed wherein antibodies specific to human cytokine polypeptide attached to a solid support and labeled human cytokine polypeptide and a sample derived from the host are passed over the solid support and the amount of label detected attached to the solid support can be correlated to a quantity of human cytokine polypeptide in the sample.

Antibodies

The polypeptides, their fragments or other derivatives, or analogs thereof, or cells expressing them can be used as an immunogen to produce antibodies thereto. These antibodies can be, for example, polyclonal or monoclonal antibodies. The present invention also includes chimeric, single chain, and humanized antibodies, as well as Fab fragments, or the product of an Fab expression library. Various procedures known in the art may be used for the production of such antibodies and fragments.

Antibodies generated against the polypeptides corresponding to a sequence of the present invention can be obtained by direct injection of the polypeptides into an animal or by administering the polypeptides to an animal, preferably a nonhuman. The antibody so obtained will then bind the polypeptides itself. In this manner, even a sequence encoding only a fragment of the polypeptides can be used to generate antibodies binding the whole native polypeptides. Such antibodies can then be used to isolate the polypeptide from tissue expressing that polypeptide.

For preparation of monoclonal antibodies, any technique which provides antibodies produced by continuous cell line cultures can be used. Examples include

the hybridoma technique (Kohler, G. and Milstein, C., Nature 256: 495-497 (1975), the trioma technique, the human B-cell hybridoma technique (Kozbor et al., Immunology Today 4: 72 (1983) and the EBV-hybridoma technique to produce human monoclonal antibodies (Cole et al., pg. 77-96 in MONOCLONAL
5 ANTIBODIES AND CANCER THERAPY, Alan R. Liss, Inc. (1985).

Techniques described for the production of single chain antibodies (U.S. Patent No. 4,946,778) can be adapted to produce single chain antibodies to immunogenic polypeptide products of this invention. Also, transgenic mice, or other organisms such as other mammals, may be used to express humanized antibodies to immunogenic
10 polypeptide products of this invention.

The above-described antibodies may be employed to isolate or to identify clones expressing the polypeptide or purify the polypeptide of the present invention by attachment of the antibody to a solid support for isolation and/or purification by affinity chromatography.

15 Thus, among others, the cytokine polypeptide of the present invention may be employed to stimulate cell proliferation and differentiation and inhibit the adhesion of PBMCs to fibronectin to treat restenosis and inflammation.

Human cytokine polypeptide binding molecules and assays

20 This invention also provides a method for identification of molecules, such as receptor molecules, that bind human cytokine polypeptide. Genes encoding proteins that bind human cytokine polypeptide, such as receptor proteins, can be identified by numerous methods known to those of skill in the art, for example, ligand panning and FACS sorting. Such methods are described in many laboratory manuals such as, for
25 instance, Coligan et al., Current Protocols in Immunology 1(2): Chapter 5 (1991).

For instance, expression cloning may be employed for this purpose. To this end polyadenylated RNA is prepared from a cell responsive to human cytokine polypeptide, a cDNA library is created from this RNA, the library is divided into pools and the pools are transfected individually into cells that are not responsive to
30 human cytokine polypeptide. The transfected cells then are exposed to labeled human

cytokine polypeptide. (human cytokine polypeptide can be labeled by a variety of well-known techniques including standard methods of radio-iodination or inclusion of a recognition site for a site-specific protein kinase.) Following exposure, the cells are fixed and binding of human cytokine polypeptide is determined. These procedures
5 conveniently are carried out on glass slides.

Pools are identified of cDNA that produced human cytokine polypeptide-binding cells. Sub-pools are prepared from these positives, transfected into host cells and screened as described above. Using an iterative sub-pooling and re-screening process, plasmids containing one or more single clones that encode the putative
10 binding molecule, such as a receptor molecule, can be isolated and the clones are sequenced.

Alternatively a labeled ligand can be photoaffinity linked to a cell extract, such as a membrane or a membrane extract, prepared from cells that express a molecule that it binds, such as a receptor molecule. Cross-linked material is resolved by
15 polyacrylamide gel electrophoresis ("PAGE") and exposed to X-ray film. The labeled complex containing the ligand-receptor can be excised, resolved into peptide fragments, and subjected to protein microsequencing. The amino acid sequence obtained from microsequencing can be used to design unique or degenerate oligonucleotide probes to screen cDNA libraries to identify genes encoding the
20 putative receptor molecule.

Polypeptides of the invention also can be used to assess human cytokine polypeptide binding capacity of human cytokine polypeptide binding molecules, such as receptor molecules, in cells or in cell-free preparations.

25 Agonists and antagonists - assays and molecules

The invention also provides a method of screening compounds to identify those which enhance or block the action of human cytokine polypeptide on cells, such as its interaction with human cytokine polypeptide-binding molecules such as receptor molecules. An agonist is a compound which increases the natural biological

functions of human cytokine polypeptide or which functions in a manner similar to human cytokine polypeptide, while antagonists decrease or eliminate such functions.

For example, a cellular compartment, such as a membrane or a preparation thereof, such as a membrane-preparation, may be prepared from a cell that expresses a molecule that binds human cytokine polypeptide, such as a molecule of a signaling or regulatory pathway modulated by human cytokine polypeptide. The preparation is incubated with labeled human cytokine polypeptide in the absence or the presence of a candidate molecule which may be a human cytokine polypeptide agonist or antagonist. The ability of the candidate molecule to bind the binding molecule is reflected in decreased binding of the labeled ligand. Molecules which bind gratuitously, i.e., without inducing the effects of human cytokine polypeptide on binding the human cytokine polypeptide binding molecule, are most likely to be good antagonists. Molecules that bind well and elicit effects that are the same as or closely related to human cytokine polypeptide, are good agonists.

Human cytokine polypeptide-like effects of potential agonists and antagonists may be measured, for instance, by determining activity of a second messenger system following interaction of the candidate molecule with a cell or appropriate cell preparation, and comparing the effect with that of human cytokine polypeptide or molecules that elicit the same effects as human cytokine polypeptide. Second messenger systems that may be useful in this regard include but are not limited to AMP guanylate cyclase, ion channel or phosphoinositide hydrolysis second messenger systems.

Another example of an assay for human cytokine polypeptide antagonists is a competitive assay that combines human cytokine polypeptide and a potential antagonist with membrane-bound human cytokine polypeptide receptor molecules or recombinant human cytokine polypeptide receptor molecules under appropriate conditions for a competitive inhibition assay. human cytokine polypeptide can be labeled, such as by radioactivity, such that the number of human cytokine polypeptide molecules bound to a receptor molecule can be determined accurately to assess the effectiveness of the potential antagonist.

Potential antagonists include small organic molecules, peptides, polypeptides and antibodies that bind to a polypeptide of the invention and thereby inhibit or extinguish its activity. Potential antagonists also may be small organic molecules, a peptide, a polypeptide such as a closely related protein or antibody that binds the same sites on a binding molecule, such as a receptor molecule, without inducing human cytokine polypeptide-induced activities, thereby preventing the action of human cytokine polypeptide by excluding human cytokine polypeptide from binding.

Other potential antagonists include antisense molecules. Antisense technology can be used to control gene expression through antisense DNA or RNA or through triple-helix formation. Antisense techniques are discussed, for example, in - Okano, J. Neurochem. 56: 560 (1991); OLIGODEOXYNUCLEOTIDES AS ANTISENSE INHIBITORS OF GENE EXPRESSION, CRC Press, Boca Raton, FL (1988). Triple helix formation is discussed in, for instance Lee et al., Nucleic Acids Research 6: 3073 (1979); Cooney et al., Science 241: 456 (1988); and Dervan et al., Science 251: 1360 (1991). The methods are based on binding of a polynucleotide to a complementary DNA or RNA. For example, the 5' coding portion of a polynucleotide that encodes the mature polypeptide of the present invention may be used to design an antisense RNA oligonucleotide of from about 10 to 40 base pairs in length. A DNA oligonucleotide is designed to be complementary to a region of the gene (or promotor) involved in transcription thereby preventing transcription and the production of human cytokine polypeptide. The antisense RNA oligonucleotide hybridizes to the mRNA in vivo and blocks translation of the mRNA molecule into human cytokine polypeptide. The oligonucleotides described above can also be delivered to cells such that the antisense RNA or DNA may be expressed in vivo to inhibit production of human cytokine polypeptide.

The antagonists may be employed in a composition with a pharmaceutically acceptable carrier, e.g., as hereinafter described.

The antagonists may be employed for instance to treat and/or prevent abnormal cellular proliferation for example, neoplasia.

Compositions

The invention also relates to compositions comprising the polynucleotide or the polypeptides discussed above or the agonists or antagonists. Thus, the polypeptides of the present invention may be employed in combination with a non-sterile or sterile carrier or carriers for use with cells, tissues or organisms, such as a pharmaceutical carrier suitable for administration to a subject. Such compositions comprise, for instance, a media additive or a therapeutically effective amount of a polypeptide of the invention and a pharmaceutically acceptable carrier or excipient. Such carriers may include, but are not limited to, saline, buffered saline, dextrose, water, glycerol, ethanol and combinations thereof. The formulation should suit the mode of administration.

Kits

The invention further relates to pharmaceutical packs and kits comprising one or more containers filled with one or more of the ingredients of the aforementioned compositions of the invention. Associated with such container(s) can be a notice in the form prescribed by a governmental agency regulating the manufacture, use or sale of pharmaceuticals or biological products, reflecting approval by the agency of the manufacture, use or sale of the product for human administration.

Administration

Polypeptides and other compounds of the present invention may be employed alone or in conjunction with other compounds, such as therapeutic compounds.

The pharmaceutical compositions may be administered in any effective, convenient manner including, for instance, administration by topical, oral, anal, vaginal, intravenous, intraperitoneal, intramuscular, subcutaneous, intranasal or intradermal routes among others.

The pharmaceutical compositions generally are administered in an amount effective for treatment or prophylaxis of a specific indication or indications. In general, the compositions are administered in an amount of at least about 10 g/kg

body weight. In most cases they will be administered in an amount not in excess of about 8 mg/kg body weight per day. Preferably, in most cases, dose is from about 10 g/kg to about 1 mg/kg body weight, daily. It will be appreciated that optimum dosage will be determined by standard methods for each treatment modality and indication, taking into account the indication, its severity, route of administration, complicating conditions and the like.

Gene therapy

The human cytokine polypeptide polynucleotides, polypeptides, agonists and antagonists that are polypeptides may be employed in accordance with the present invention by expression of such polypeptides in vivo, in treatment modalities often referred to as "gene therapy."

Thus, for example, cells from a patient may be engineered with a polynucleotide, such as a DNA or RNA, encoding a polypeptide ex vivo, and the engineered cells then can be provided to a patient to be treated with the polypeptide. For example, cells may be engineered ex vivo by the use of a retroviral plasmid vector containing RNA encoding a polypeptide of the present invention. Such methods are well-known in the art and their use in the present invention will be apparent from the teachings herein.

Similarly, cells may be engineered in vivo for expression of a polypeptide in vivo by procedures known in the art. For example, a polynucleotide of the invention may be engineered for expression in a replication defective retroviral vector, as discussed above. The retroviral expression construct then may be isolated and introduced into a packaging cell is transduced with a retroviral plasmid vector containing RNA encoding a polypeptide of the present invention such that the packaging cell now produces infectious viral particles containing the gene of interest. These producer cells may be administered to a patient for engineering cells in vivo and expression of the polypeptide in vivo. These and other methods for administering a polypeptide of the present invention by such method should be apparent to those skilled in the art from the teachings of the present invention.

Retroviruses from which the retroviral plasmid vectors herein above mentioned may be derived include, but are not limited to, Moloney Murine Leukemia Virus, spleen necrosis virus, retroviruses such as Rous Sarcoma Virus, Harvey Sarcoma Virus, avian leukosis virus, gibbon ape leukemia virus, human immunodeficiency virus, 5 adenovirus, Myeloproliferative Sarcoma Virus, and mammary tumor virus. In one embodiment, the retroviral plasmid vector is derived from Moloney Murine Leukemia Virus.

Such vectors well include one or more promoters for expressing the polypeptide. Suitable promoters which may be employed include, but are not limited 10 to, the retroviral LTR; the SV40 promoter; and the human cytomegalovirus (CMV) promoter described in Miller et al., Biotechniques 7: 980-990 (1989), or any other promoter (e.g., cellular promoters such as eukaryotic cellular promoters including, but not limited to, the histone, RNA polymerase III, and α -actin promoters). Other viral promoters which may be employed include, but are not limited to, adenovirus 15 promoters, thymidine kinase (TK) promoters, and B19 parvovirus promoters. The selection of a suitable promoter will be apparent to those skilled in the art from the teachings contained herein.

The nucleic acid sequence encoding the polypeptide of the present invention will be placed under the control of a suitable promoter. Suitable promoters which 20 may be employed include, but are not limited to, adenoviral promoters, such as the adenoviral major late promoter; or heterologous promoters, such as the cytomegalovirus (CMV) promoter; the respiratory syncytial virus (RSV) promoter; inducible promoters, such as the MMT promoter, the metallothionein promoter; heat shock promoters; the albumin promoter; the ApoAI promoter; human globin 25 promoters; viral thymidine kinase promoters, such as the Herpes Simplex thymidine kinase promoter; retroviral LTRs (including the modified retroviral LTRs herein above described); the α -actin promoter; and human growth hormone promoters. The promoter also may be the native promoter which controls the gene encoding the polypeptide.

The retroviral plasmid vector is employed to transduce packaging cell lines to form producer cell lines. Examples of packaging cells which may be transfected include, but are not limited to, the PE501, PA317, Y-2, Y-AM, PA12, T19-14X, VT-19-17-H2, YCRE, YCRIP, GP+E-86, GP+envAm12, and DAN cell lines as described in Miller, A., Human Gene Therapy 1: 5-14 (1990). The vector may be transduced into the packaging cells through any means known in the art. Such means include, but are not limited to, electroporation, the use of liposomes, and CaPO4 precipitation. In one alternative, the retroviral plasmid vector may be encapsulated into a liposome, or coupled to a lipid, and then administered to a host.

10 The producer cell line will generate infectious retroviral vector particles, which include the nucleic acid sequence(s) encoding the polypeptides. Such retroviral vector particles then may be employed to transduce eukaryotic cells, either in vitro or in vivo. The transduced eukaryotic cells will express the nucleic acid sequence(s) encoding the polypeptide. Eukaryotic cells which may be transduced include, but are not limited to, embryonic stem cells, embryonic carcinoma cells, as well as hematopoietic stem cells, hepatocytes, fibroblasts, myoblasts, keratinocytes, endothelial cells, and bronchial epithelial cells.

EXAMPLES

The present invention is further described by the following examples. The examples are provided solely to illustrate the invention by reference to specific embodiments. These exemplification's, while illustrating certain specific aspects of the invention, do not portray the limitations or circumscribe the scope of the disclosed invention.

Certain terms used herein are explained in the foregoing glossary.

All examples were carried out using standard techniques, which are well known and routine to those of skill in the art, except where otherwise described in detail.

10 Routine molecular biology techniques of the following examples can be carried out as described in standard laboratory manuals, such as Sambrook et al., MOLECULAR CLONING: A LABORATORY MANUAL, 2nd Ed.; Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989), herein referred to as "Sambrook."

All parts or amounts set out in the following examples are by weight, unless otherwise specified.

Unless otherwise stated size separation of fragments in the examples below was carried out using standard techniques of agarose and polyacrylamide gel electrophoresis ("PAGE") in Sambrook and numerous other references such as, for instance, by Goeddel et al., Nucleic Acids Res. 8: 4057 (1980).

20 Unless described otherwise, ligations were accomplished using standard buffers, incubation temperatures and times, approximately equimolar amounts of the DNA fragments to be ligated and approximately 10 units of T4 DNA ligase ("ligase") per 0.5 g of DNA.

25 Example 1 Expression and purification of mature human cytokine polypeptide using bacteria

The DNA sequence encoding human cytokine polypeptide in the deposited polynucleotide was amplified using PCR oligonucleotide primers specific to the amino acid carboxyl terminal sequence of the human cytokine polypeptide protein and to vector sequences 3' to the gene. Additional nucleotides containing restriction sites

to facilitate cloning were added to the 5' and 3' sequences respectively. The 5' oligonucleotide primer had the sequence 5' GTAC

GGATCCAGCTGTAACAAAGCACTCTGTG 3' (SEQ ID NO:3) containing the underlined Bam HI restriction site, which encodes a start AUG, followed by 22 nucleotides of the human cytokine polypeptide coding sequence set out in Figure 1 (SEQ ID NO:1) beginning with the second base of the twenty-first codon.

The 3' primer had the sequence 5' GTCAAGCTTTTAA AACATGCAGTTCATACATTTGAC 3' (SEQ ID NO:4) containing the underlined Hind III restriction site, 6-His tag, followed by 27 nucleotides complementary to the last 27 nucleotides of the human cytokine polypeptide coding sequence set out in Figure 1 (SEQ ID NO:1), including the stop codon.

The restriction sites were convenient to restriction enzyme sites in the bacterial expression vectors pQE-9, which were used for bacterial expression in these examples. (Qiagen, Inc. Chatsworth, CA). pQE-9 encodes ampicillin antibiotic resistance ("Ampr") and contains a bacterial origin of replication ("ori"), an IPTG inducible promoter, a ribosome binding site ("RBS"), a 6-His tag and restriction enzyme sites.

The amplified human cytokine polypeptide DNA and the vector pQE-9 both were digested with Bam HI and Hind III and the digested DNAs then were ligated together. Insertion of the human cytokine polypeptide DNA into the restricted vector placed the human cytokine polypeptide coding region downstream of and operably linked to the vector's IPTG-inducible promoter and in-frame with an initiating AUG appropriately positioned for translation of human cytokine polypeptide.

The ligation mixture was transformed into competent *E. coli* cells using standard procedures. Such procedures are described in Sambrook et al., MOLECULAR CLONING: A LABORATORY MANUAL, 2nd Ed.; Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989). *E. coli* strain M15/rep4, containing multiple copies of the plasmid pREP4, which expresses lac repressor and confers kanamycin resistance ("Kanr"), was used in carrying out the illustrative

example described here. This strain, which is only one of many that are suitable for expressing human cytokine polypeptide, is available commercially from Qiagen.

Transformants were identified by their ability to grow on LB plates in the presence of ampicillin. Plasmid DNA was isolated from resistant colonies and the
5 identity of the cloned DNA was confirmed by restriction analysis.

Clones containing the desired constructs were grown overnight ("O/N") in liquid culture in LB media supplemented with both ampicillin (100 ug/ml) and kanamycin (25 ug/ml).

The O/N culture was used to inoculate a large culture, at a dilution of
10 approximately 1:100 to 1:250. The cells were grown to an optical density at 600nm ("OD600") of between 0.4 and 0.6. Isopropyl-B-D-thiogalactopyranoside ("IPTG") was then added to a final concentration of 1 mM to induce transcription from lac repressor sensitive promoters, by inactivating the lacI repressor. Cells subsequently were incubated further for 3 to 4 hours. Cells then were harvested by centrifugation
15 and disrupted, by standard methods. Inclusion bodies were purified from the disrupted cells using routine collection techniques, and protein was solubilized from the inclusion bodies into 8M urea. The 8M urea solution containing the solubilized protein was passed over a PD-10 column in 2X phosphate buffered saline ("PBS"), thereby removing the urea, exchanging the buffer and refolding the protein. The
20 protein was purified by a further step of chromatography to remove endotoxin. Then, it was sterile filtered. The sterile filtered protein preparation was stored in 2X PBS at a concentration of 95 micrograms per mL.

Analysis of the preparation by standard methods of polyacrylamide gel electrophoresis revealed that the preparation had the expected molecular weight of,
25 approximately, 26 kDa.

Example 2 Cloning and expression of human cytokine polypeptide in a baculovirus expression system

The cDNA sequence encoding the full length human cytokine polypeptide protein, in the deposited clone is amplified using PCR oligonucleotide primers corresponding to the 5' and 3' sequences of the gene:

The 5' primer has the sequence 5' GATAGGATCCGCC
5 ATCATGAAGTTACACT ATGTTGCTGTG 3' (SEQ ID NO:5) containing the underlined Bam HI restriction enzyme site followed by 24 bases of the sequence of human cytokine polypeptide of Figure 1 (SEQ ID NO:2). Inserted into an expression vector, as described below, the 5' end of the amplified fragment encoding human cytokine polypeptide provides an efficient signal peptide. An efficient signal for
10 initiation of translation in eukaryotic cells, as described by Kozak, M., J. Mol. Biol. 196: 947-950 (1987) is appropriately located in the primer portion of the construct.

The 3' primer has the sequence 5' GTCAGGTACCTTA
AAACATGCAGTTCATACATTTGAC 3' (SEQ ID NO:6) containing the underlined KpnI restriction followed by nucleotides complementary to the last 27
15 nucleotides of the human cytokine polypeptide coding sequence set out in Figure 1 (SEQ ID NO:1), including the stop codon.

The amplified fragment is isolated from a 1% agarose gel using a commercially available kit ("GeneClean," BIO 101 Inc., La Jolla, Ca.). The fragment then is digested with BamH1 and Asp718 and again is purified on a 1% agarose gel. This fragment is
20 designated herein F2.

The vector pA2 is used to express the human cytokine polypeptide protein in the baculovirus expression system, using standard methods, such as those described in Summers et al, A MANUAL OF METHODS FOR BACULOVIRUS VECTORS AND INSECT CELL CULTURE PROCEDURES, Texas Agricultural Experimental
25 Station Bulletin No. 1555 (1987). This expression vector contains the strong polyhedrin promoter of the Autographa californica nuclear polyhedrosis virus (AcMNPV) followed by convenient restriction sites. The signal peptide of AcMNPV gp67, including the N-terminal methionine, is located just upstream of a BamH1 site. The polyadenylation site of the simian virus 40 ("SV40") is used for efficient
30 polyadenylation. For an easy selection of recombinant virus the beta-galactosidase

gene from E.coli is inserted in the same orientation as the polyhedrin promoter and is followed by the polyadenylation signal of the polyhedrin gene. The polyhedrin sequences are flanked at both sides by viral sequences for cell-mediated homologous recombination with wild-type viral DNA to generate viable virus that express the cloned polynucleotide.

Many other baculovirus vectors could be used in place of pA2, such as pAc373, pVL941 and pAcIM1 provided, as those of skill readily will appreciate, that construction provides appropriately located signals for transcription, translation, trafficking and the like, such as an in-frame AUG and a signal peptide, as required.

Such vectors are described in Luckow et al., Virology 170: 31-39, among others.

The plasmid is digested with the restriction enzymes Bam HI and KpnI and then is dephosphorylated using calf intestinal phosphatase, using routine procedures known in the art. The DNA is then isolated from a 1% agarose gel using a commercially available kit ("GeneClean" BIO 101 Inc., La Jolla, Ca.). This vector

DNA is designated herein "V2".

Fragment F2 and the dephosphorylated plasmid V2 are ligated together with T4 DNA ligase. E.coli HB101 cells are transformed with ligation mix and spread on culture plates. Bacteria are identified that contain the plasmid with the human cytokine polypeptide gene by digesting DNA from individual colonies using Bam HI and KpnI and then analyzing the digestion product by gel electrophoresis. The sequence of the cloned fragment is confirmed by DNA sequencing. This plasmid is designated herein pBac human cytokine polypeptide.

5 g of the plasmid pBac human cytokine polypeptide is co-transfected with 1.0 g of a commercially available linearized baculovirus DNA ("BaculoGold baculovirus DNA", Pharmingen, San Diego, CA.), using the lipofection method described by Felgner et al., Proc. Natl. Acad. Sci. USA 84: 7413-7417 (1987). 1 g of BaculoGold virus DNA and 5 g of the plasmid pBac human cytokine polypeptide are mixed in a sterile well of a microtiter plate containing 50 l of serum free Grace's medium (Life Technologies Inc., Gaithersburg, MD). Afterwards 10 l Lipofectin plus 90 l Grace's medium are added, mixed and incubated for 15 minutes at room

temperature. Then the transfection mixture is added drop-wise to Sf9 insect cells (ATCC CRL 1711) seeded in a 35 mm tissue culture plate with 1 ml Grace's medium without serum. The plate is rocked back and forth to mix the newly added solution. The plate is then incubated for 5 hours at 27 C. After 5 hours the transfection solution is removed from the plate and 1 ml of Grace's insect medium supplemented with 10% fetal calf serum is added. The plate is put back into an incubator and cultivation is continued at 27 C for four days.

After four days the supernatant is collected and a plaque assay is performed, as described by Summers and Smith, cited above. An agarose gel with "Blue Gal" (Life Technologies Inc., Gaithersburg) is used to allow easy identification and isolation of gal-expressing clones, which produce blue-stained plaques. (A detailed description of a "plaque assay" of this type can also be found in the user's guide for insect cell culture and baculovirology distributed by Life Technologies Inc., Gaithersburg, page 9-10).

Four days after serial dilution, the virus is added to the cells. After appropriate incubation, blue stained plaques are picked with the tip of an Eppendorf pipette. The agar containing the recombinant viruses is then resuspended in an Eppendorf tube containing 200 μ l of Grace's medium. The agar is removed by a brief centrifugation and the supernatant containing the recombinant baculovirus is used to infect Sf9 cells seeded in 35 mm dishes. Four days later the supernatants of these culture dishes are harvested and then they are stored at 4 C. A clone containing properly inserted human cytokine polypeptide is identified by DNA analysis including restriction mapping and sequencing. This is designated herein as V-human cytokine polypeptide.

Sf9 cells are grown in Grace's medium supplemented with 10% heat-inactivated FBS. The cells are infected with the recombinant baculovirus V-human cytokine polypeptide at a multiplicity of infection ("MOI") of about 2 (about 1 to about 3). Six hours later the medium is removed and is replaced with SF900 II medium minus methionine and cysteine (available from Life Technologies Inc., Gaithersburg). 42 hours later, 5 μ Ci of 35S-methionine and 5 μ Ci 35S cysteine

(available from Amersham) are added. The cells are further incubated for 16 hours and then they are harvested by centrifugation, lysed and the labeled proteins are visualized by SDS-PAGE and autoradiography.

5 Example 3 Expression of human cytokine polypeptide in COS cells

The expression plasmid, human cytokine polypeptide HA, is made by cloning a cDNA encoding human cytokine polypeptide into the expression vector pcDNA1/Amp (which can be obtained from Invitrogen, Inc.).

- 10 The expression vector pcDNA1/amp contains: (1) an E.coli origin of replication effective for propagation in E. coli and other prokaryotic cell; (2) an ampicillin resistance gene for selection of plasmid-containing prokaryotic cells; (3) an SV40 origin of replication for propagation in eukaryotic cells; (4) a CMV promoter, a polylinker, an SV40 intron, and a polyadenylation signal arranged so that a cDNA
- 15 conveniently can be placed under expression control of the CMV promoter and operably linked to the SV40 intron and the polyadenylation signal by means of restriction sites in the polylinker.

- A DNA fragment encoding the entire human cytokine polypeptide precursor and a HA tag fused in frame to its 3' end is cloned into the polylinker region of the
- 20 vector so that recombinant protein expression is directed by the CMV promoter. The HA tag corresponds to an epitope derived from the influenza hemagglutinin protein described by Wilson et al., Cell 37: 767 (1984). The fusion of the HA tag to the target protein allows easy detection of the recombinant protein with an antibody that recognizes the HA epitope.

- 25 The plasmid construction strategy is as follows.

The human cytokine polypeptide cDNA of the deposit clone is amplified using primers that contained convenient restriction sites, much as described above regarding the construction of expression vectors for expression of human cytokine polypeptide in E. coli and S. furgiperda.

To facilitate detection, purification and characterization of the expressed human cytokine polypeptide, one of the primers contains a hemagglutinin tag ("HA tag") as described above.

Suitable primers include that following, which are used in this example.

- 5 The 5' primer, containing the underlined BamHI site, an AUG start codon, hemagglutinin tag, and 24 bps of human cytokine polypeptide sequence, has the following sequence, 5'

GATAGGATCCGCCATCATGAGCGTAGTCTGGGACGTCGTATGGGTAAA
GTTACACTATGTTGCTGTG 3' (SEQ ID NO:7).

- 10 The 3' primer, containing the underlined KpnI site and 27 bp of 3' coding sequence (at the 3' end) has the following sequence, 5'
GTCAGGTACCTTAAAACATGCAGTTCATACATTGAC 3' (SEQ ID NO:8).

- 15 The PCR amplified DNA fragment and the vector, pcDNAI/Amp, are digested with and then ligated. The ligation mixture is transformed into E. coli strain SURE (Stratagene Cloning Systems, La Jolla, CA) the transformed culture is plated on ampicillin media plates which then are incubated to allow growth of ampicillin resistant colonies. Plasmid DNA is isolated from resistant colonies and examined by restriction analysis and gel sizing for the presence of the human cytokine polypeptide-encoding fragment.

- 20 For expression of recombinant human cytokine polypeptide, COS cells are transfected with an expression vector, as described above, using DEAE-DEXTRAN, as described, for instance, in Sambrook et al., MOLECULAR CLONING: A LABORATORY MANUAL, Cold Spring Laboratory Press, Cold Spring Harbor, New York (1989).

- 25 Cells are incubated under conditions for expression of human cytokine polypeptide by the vector.

- 30 Expression of the human cytokine polypeptide HA fusion protein is detected by radiolabelling and immunoprecipitation, using methods described in, for example Harlow et al., ANTIBODIES: A LABORATORY MANUAL, 2nd Ed.; Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (1988). To this end, two

days after transfection, the cells are labeled by incubation in media containing ³⁵S-cysteine for 8 hours. The cells and the media are collected, and the cells are washed and the lysed with detergent-containing RIPA buffer: 150 mM NaCl, 1% NP-40, 0.1% SDS, 1% NP-40, 0.5% DOC, 50 mM TRIS, pH 7.5, as described by Wilson et al. cited above. Proteins are precipitated from the cell lysate and from the culture media using an HA-specific monoclonal antibody. The precipitated proteins then are analyzed by SDS-PAGE gels and autoradiography. An expression product of the expected size is seen in the cell lysate, which is not seen in negative controls.

10 Example 4 Tissue distribution of human cytokine polypeptide expression

Northern blot analysis is carried out to examine the levels of expression of human cytokine polypeptide in human tissues, using methods described by, among others, Sambrook et al, cited above. Total cellular RNA samples are isolated with RNAzol B system (Biotech Laboratories, Inc. 6023 South Loop East, Houston, TX 77033).

About 10 g of Total RNA is isolated from tissue samples. The RNA is size resolved by electrophoresis through a 1% agarose gel under strongly denaturing conditions. RNA is blotted from the gel onto a nylon filter, and the filter then is prepared for hybridization to a detectably labeled polynucleotide probe.

As a probe to detect mRNA that encodes human cytokine polypeptide, the antisense strand of the coding region of the cDNA insert in the deposited clone is labeled to a high specific activity. The cDNA is labeled by primer extension, using the Prime-It kit, available from Stratagene. The reaction is carried out using 50 ng of the cDNA, following the standard reaction protocol as recommended by the supplier. The labeled polynucleotide is purified away from other labeled reaction components by column chromatography using a Select-G-50 column, obtained from 5-Prime - 3-Prime, Inc. of 5603 Arapahoe Road, Boulder, CO 80303.

The labeled probe is hybridized to the filter, at a concentration of 1,000,000 cpm/ml, in a small volume of 7% SDS, 0.5 M NaPO₄, pH 7.4 at 65 C, overnight.

Thereafter the probe solution is drained and the filter is washed twice at room temperature and twice at 60 C with 0.5 x SSC, 0.1% SDS. The filter then is dried and exposed to film at -70 C overnight with an intensifying screen.

5 Autoradiography shows that mRNA for human cytokine polypeptide is abundant in most major tissues including heart, brain, lung, placenta, kidney, skeletal muscle, ovary, testis, prostate, and smooth muscle.

Example 5 Gene therapeutic expression of human cytokine polypeptide

10 Fibroblasts are obtained from a subject by skin biopsy. The resulting tissue is placed in tissue-culture medium and separated into small pieces. Small chunks of the tissue are placed on a wet surface of a tissue culture flask, approximately ten pieces are placed in each flask. The flask is turned upside down, closed tight and left at room temperature overnight. After 24 hours at room temperature, the flask is inverted - the chunks of tissue remain fixed to the bottom of the flask - and fresh media is added
15 (e.g., Ham's F12 media, with 10% FBS, penicillin and streptomycin). The tissue is then incubated at 37 C for approximately one week. At this time, fresh media is added and subsequently changed every several days. After an additional two weeks in culture, a monolayer of fibroblasts emerges. The monolayer is trypsinized and scaled into larger flasks.

20 A vector for gene therapy is digested with restriction enzymes for cloning a fragment to be expressed. The digested vector is treated with calf intestinal phosphatase to prevent self-ligation. The dephosphorylated, linear vector is fractionated on an agarose gel and purified.

human cytokine polypeptide cDNA capable of expressing active human
25 cytokine polypeptide, is isolated. The ends of the fragment are modified, if necessary, for cloning into the vector. For instance, 5' overhanging may be treated with DNA polymerase to create blunt ends. 3' overhanging ends may be removed using S1 nuclease. Linkers may be ligated to blunt ends with T4 DNA ligase.

30 Equal quantities of the Moloney murine leukemia virus linear backbone and the human cytokine polypeptide fragment are mixed together and joined using T4 DNA

ligase. The ligation mixture is used to transform E. Coli and the bacteria are then plated onto agar-containing kanamycin. Kanamycin phenotype and restriction analysis confirm that the vector has the properly inserted gene.

5 Packaging cells are grown in tissue culture to confluent density in Dulbecco's Modified Eagles Medium (DMEM) with 10% calf serum (CS), penicillin and streptomycin. The vector containing the human cytokine polypeptide gene is introduced into the packaging cells by standard techniques. Infectious viral particles containing the human cytokine polypeptide gene are collected from the packaging cells, which now are called producer cells.

10 Fresh media is added to the producer cells, and after an appropriate incubation period media is harvested from the plates of confluent producer cells. The media, containing the infectious viral particles, is filtered through a Millipore filter to remove detached producer cells. The filtered media then is used to infect fibroblast cells. Media is removed from a sub-confluent plate of fibroblasts and quickly replaced with
15 the filtered media. Polybrene (Aldrich) may be included in the media to facilitate transduction. After appropriate incubation, the media is removed and replaced with fresh media. If the titer of virus is high, then virtually all fibroblasts will be infected and no selection is required. If the titer is low, then it is necessary to use a retroviral vector that has a selectable marker, such as neo or his, to select out transduced cells
20 for expansion.

Engineered fibroblasts then may be injected into rats, either alone or after having been grown to confluence on microcarrier beads, such as cytodex 3 beads. The injected fibroblasts produce human cytokine polypeptide product, and the biological actions of the protein are conveyed to the host.

25 It will be clear that the invention may be practiced otherwise than as particularly described in the foregoing description and examples.

Numerous modifications and variations of the present invention are possible in light of the above teachings and, therefore, are within the scope of the appended claims.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: LI, YI
SU, KUI
LI, HAODONG
- (ii) TITLE OF INVENTION: HUMAN CYTOKINE POLYPEPTIDE
- (iii) NUMBER OF SEQUENCES: 9
- (iv) CORRESPONDENCE ADDRESS:
(A) ADDRESSEE: HUMAN GENOME SCIENCES, INC.
(B) STREET: 9410 KEY WEST AVENUE
(C) CITY: ROCKVILLE
(D) STATE: MARYLAND
(E) COUNTRY: USA
(F) ZIP: 20850
- (v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER: US
(B) FILING DATE: 20-MAR-1997
(C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
(A) NAME: Brookes, Anders, A.
(B) REGISTRATION NUMBER: 36,373
(C) REFERENCE/DOCKET NUMBER: PF270
- (ix) TELECOMMUNICATION INFORMATION:
(A) TELEPHONE: 301-309-8504
(B) TELEFAX: 301-309-8512

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 780 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:
(A) NAME/KEY: sig_peptide

(B) LOCATION: 72..131

(ix) FEATURE:

(A) NAME/KEY: mat_peptide

(B) LOCATION: 132..740

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 72..740

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

GCACGAGGGC GACTTCGCGG GACCGTGGCG CATGTTTCCT GGGAGTTACT GATCATCTTC	60
TTTGAAGAAA C ATG AAG TTA CAC TAT GTT GCT GTG CTT ACT CTA GCC ATC	110
Met Lys Leu His Tyr Val Ala Val Leu Thr Leu Ala Ile	
-20 -15 -10	
CTG ATG TTC CTG ACA TGG CTT CCA GAA TCA CTG AGC TGT AAC AAA GCA	158
Leu Met Phe Leu Thr Trp Leu Pro Glu Ser Leu Ser Cys Asn Lys Ala	
-5 1 5	
CTC TGT GCT AGT GAT GTG AGC AAA TGC CTC ATT CAG GAG CTC TGC CAG	206
Leu Cys Ala Ser Asp Val Ser Lys Cys Leu Ile Gln Glu Leu Cys Gln	
10 15 20 25	
TGC CGG CCG GGA GAA GGC AAT TGC TCC TGC TGT AAG GAG TGC ATG CTG	254
Cys Arg Pro Gly Glu Gly Asn Cys Ser Cys Cys Lys Glu Cys Met Leu	
30 35 40	
TGT CTT GGG GCC CTT TGG GAC GAG TGC TGT GAC TGT GTT GGT ATG TGT	302
Cys Leu Gly Ala Leu Trp Asp Glu Cys Cys Asp Cys Val Gly Met Cys	
45 50 55	
AAT CCT CGA AAT TAT AGT GAC ACA CCT CCA ACT TCA AAG AGC ACA GTG	350
Asn Pro Arg Asn Tyr Ser Asp Thr Pro Pro Thr Ser Lys Ser Thr Val	
60 65 70	
GAG GAG CTG CAT GAA CCG ATC CCT TCT CTC TTC CGG GCA CTC ACA GAA	398
Glu Glu Leu His Glu Pro Ile Pro Ser Leu Phe Arg Ala Leu Thr Glu	
75 80 85	
GGA GAT ACT CAG TTG AAT TGG AAC ATC GTT TCT TTC CCT GTT GCA GAA	446
Gly Asp Thr Gln Leu Asn Trp Asn Ile Val Ser Phe Pro Val Ala Glu	
90 95 100 105	
GAA CTT TCA CAT CAT GAG AAT CTG GTT TCA TTT TTA GAA ACT GTG AAC	494
Glu Leu Ser His His Glu Asn Leu Val Ser Phe Leu Glu Thr Val Asn	
110 115 120	
CAG CCA CAC CAC CAG AAT GTG TCT GTC CCC AGC AAT AAT GTT CAC GCG	542
Gln Pro His His Gln Asn Val Ser Val Pro Ser Asn Asn Val His Ala	
125 130 135	

CCT TAT TCC AGT GAC AAA GAA CAC ATG TGT ACT GTG GTT TAT TTT GAT	590
Pro Tyr Ser Ser Asp Lys Glu His Met Cys Thr Val Val Tyr Phe Asp	
140 145 150	
GAC TGC ATG TCC ATA CAT CAG TGT AAA ATA TCC TGT GAG TCC ATG GGA	638
Asp Cys Met Ser Ile His Gln Cys Lys Ile Ser Cys Glu Ser Met Gly	
155 160 165	
GCA TCC AAA TAT CGC TGG TTT CAT AAT GCC TGC TGC GAG TGC ATT GGT	686
Ala Ser Lys Tyr Arg Trp Phe His Asn Ala Cys Cys Glu Cys Ile Gly	
170 175 180 185	
CCA GAA TGT ATT GAC TAT GGT AGT AAA ACT GTC AAA TGT ATG AAC TGC	734
Pro Glu Cys Ile Asp Tyr Gly Ser Lys Thr Val Lys Cys Met Asn Cys	
190 195 200	
ATG TTT TAAAGAAGAC AAATGCAAAC CAAAGCAACT TAGTAAATA	780
Met Phe	

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 223 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Lys Leu His Tyr Val Ala Val Leu Thr Leu Ala Ile Leu Met Phe	
-20 -15 -10 -5	
Leu Thr Trp Leu Pro Glu Ser Leu Ser Cys Asn Lys Ala Leu Cys Ala	
1 5 10	
Ser Asp Val Ser Lys Cys Leu Ile Gln Glu Leu Cys Gln Cys Arg Pro	
15 20 25	
Gly Glu Gly Asn Cys Ser Cys Cys Lys Glu Cys Met Leu Cys Leu Gly	
30 35 40	
Ala Leu Trp Asp Glu Cys Cys Asp Cys Val Gly Met Cys Asn Pro Arg	
45 50 55 60	
Asn Tyr Ser Asp Thr Pro Pro Thr Ser Lys Ser Thr Val Glu Glu Leu	
65 70 75	
His Glu Pro Ile Pro Ser Leu Phe Arg Ala Leu Thr Glu Gly Asp Thr	
80 85 90	
Gln Leu Asn Trp Asn Ile Val Ser Phe Pro Val Ala Glu Glu Leu Ser	
95 100 105	

His	His	Glu	Asn	Leu	Val	Ser	Phe	Leu	Glu	Thr	Val	Asn	Gln	Pro	His
110						115					120				
His	Gln	Asn	Val	Ser	Val	Pro	Ser	Asn	Asn	Val	His	Ala	Pro	Tyr	Ser
125					130					135					140
Ser	Asp	Lys	Glu	His	Met	Cys	Thr	Val	Val	Tyr	Phe	Asp	Asp	Cys	Met
				145					150					155	
Ser	Ile	His	Gln	Cys	Lys	Ile	Ser	Cys	Glu	Ser	Met	Gly	Ala	Ser	Lys
			160					165					170		
Tyr	Arg	Trp	Phe	His	Asn	Ala	Cys	Cys	Glu	Cys	Ile	Gly	Pro	Glu	Cys
	175						180					185			
Ile	Asp	Tyr	Gly	Ser	Lys	Thr	Val	Lys	Cys	Met	Asn	Cys	Met	Phe	
	190						195				200				

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 32 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GTACGGATCC AGCTGTAACA AAGCACTCTG TG

32

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 36 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

GTCAAGCTTT TAAACATGC AGTTCATACA TTTGAC

36

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 40 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

GATAGGATCC GCCATCATGA AGTTACACTA TGTGCTGTG

40

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 37 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

GTCAGGTACC TTAAAACATG CAGTTCATAC ATTTGAC

37

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 67 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

GATAGGATCC GCCATCATGA GCGTAGTCTG GGACGTCGTA TGGGTAAAGT TACACTATGT

60

TGCTGTG

67

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 37 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

GTCAGGTACC TTAAAACATG CAGTTCATAC ATTTGAC

37

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 206 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Met Gln Leu Leu Cys Tyr Phe Val Ile Leu Phe Val Gly Ile Ala Pro
1 5 10 15
Trp Ser Ser Leu Ala Asn Asp Asp Gly Cys Asn Glu Val Val Cys Gly
20 25 30
Ser Val Val Ser Lys Cys Leu Ile Thr Gln Ser Cys Gln Cys Lys Leu
35 40 45
Asn Asp Cys His Cys Cys Lys Asp Cys Leu Asn Cys Leu Gly Glu Leu
50 55 60
Tyr Ile Glu Cys Cys Gly Cys Leu Asp Met Cys Pro Lys His Lys Asp
65 70 75 80
Val Leu Pro Ser Leu Thr Pro Arg Ser Glu Ile Gly Asp Ile Glu Gly
85 90 95
Val Pro Glu Leu Phe Asp Thr Leu Thr Ala Glu Asp Asp Glu Gly Trp
100 105 110
Ser Thr Ile Arg Phe Ser Met Arg Ala Gly Phe Lys Gln Arg Val Ala
115 120 125
Gln Gly Gly Ala Ser Gly Asp Ala Gly Asn Gly Asn Gly Asn Gly Asn
130 135 140
Ala Gly Ser Ala Gly Val Thr Leu Cys Thr Val Ile Tyr Asn Ser Cys
145 150 155 160
Ile Arg Ala Asn Lys Cys Arg Gln Gln Cys Glu Ser Met Gly Ala Ser
165 170 175
Ser Tyr Arg Trp Phe His Asp Gly Cys Cys Glu Cys Val Gly Glu Asn
180 185 190
Cys Leu Asn Tyr Gly Ile Asn Glu Ser Arg Cys Arg Gly Cys
195 200 205

WHAT IS CLAIMED IS:

1. An isolated polynucleotide comprising a polynucleotide having at least a 70% identity to a member selected from the group consisting of:

(a) a polynucleotide encoding a polypeptide comprising an amino acid sequence as set forth in SEQ ID NO:2;

(b) a polynucleotide encoding a polypeptide comprising amino acid 1 to amino acid 203 set forth in SEQ ID NO:2

(c) a polynucleotide which is complementary to the polynucleotide of (a) or (b); and

(d) a polynucleotide comprising at least 15 bases of SEQ ID NO:1.

2. The polynucleotide of Claim 1 wherein the polynucleotide is DNA.

3. The polynucleotide of Claim 1 wherein the polynucleotide is RNA.

4. The polynucleotide of Claim 1 wherein the polynucleotide is genomic DNA.

5. The polynucleotide of Claim 2 which encodes a polypeptide comprising amino acid 1 to 203 of SEQ ID NO:2.

6. The polynucleotide of Claim 2 which encodes a polypeptide comprising the amino acid sequence as set forth in SEQ ID NO:2.

7. An isolated polynucleotide comprising a polynucleotide having at least a 70% identity to a member selected from the group consisting of:

(a) a polynucleotide which encodes a mature polypeptide having the amino acid sequence expressed by the human cDNA contained in the deposited clone;

(b) a polynucleotide which is complementary to the polynucleotide of (a); and

(c) a polynucleotide comprising at least 15 bases of the polynucleotide of (a) or (b).

8. The polynucleotide of claim 1 comprising the sequence as set forth in SEQ ID No. 1 from nucleotide 1 to nucleotide 780.

9. The polynucleotide of claim 1 comprising the sequence as set forth in SEQ ID No. 1 from nucleotide 132 to nucleotide 780.

10. A vector comprising the DNA of Claim 2.

11. A host cell comprising the vector of Claim 10.

12. A process for producing a polypeptide comprising: expressing from the host cell of Claim 11 the polypeptide encoded by said DNA.

13. A process for producing a cell which expresses a polypeptide comprising genetically engineering the cell with the vector of Claim 10.

14. A polypeptide comprising a member selected from the group consisting of:

(a) a polypeptide having an amino acid sequence set forth in SEQ ID NO:2;
and

(b) a polypeptide which is at least 70% identical to the polypeptide of (a).

15. The polypeptide of Claim 14 wherein the polypeptide comprises an amino acid sequence as set forth in SEQ ID NO:2.

16. The polypeptide of Claim 14 wherein the polypeptide comprises amino acid 1 to amino acid 203 of SEQ ID NO:2.

17. A compound which inhibits activation of the polypeptide of claim 14.
18. A compound which activates the polypeptide of claim 14.
19. A method for the treatment of a patient having need of human cytokine polypeptide comprising: administering to the patient a therapeutically effective amount of the polypeptide of claim 14.
20. The method of Claim 19 wherein said therapeutically effective amount of the polypeptide is administered by providing to the patient DNA encoding said polypeptide and expressing said polypeptide in vivo.
21. A method for the treatment of a patient having need to inhibit a human cytokine polypeptide comprising: administering to the patient a therapeutically effective amount of the compound of Claim 17.
22. A process for diagnosing a disease or a susceptibility to a disease related to an under-expression of the polypeptide of claim 14 comprising:
determining a mutation in a nucleic acid sequence encoding said polypeptide.
23. A diagnostic process comprising:
analyzing for the presence of the polypeptide of claim 14 in a sample derived from a host.
24. A method for identifying compounds which bind to and inhibit activation of the polypeptide of claim 14 comprising: contacting a cell expressing on the surface thereof a receptor for the polypeptide, said receptor being associated with a second component capable of providing a detectable signal in response to the binding of a compound to said receptor, with an analytically detectable human cytokine polypeptide and a compound under conditions to permit binding to the receptor; and

ABSTRACT OF THE DISCLOSURE

The invention relates to human cytokine polypeptides, polynucleotides encoding the polypeptides, methods for producing the polypeptides, in particular by expressing the polynucleotides, and agonists and antagonists of the polypeptides. The invention further relates to methods for utilizing such polynucleotides, polypeptides, agonists and antagonists for applications, which relate, in part, to research, diagnostic and clinical arts.

10 30 50
 GCACGAGGGCGACTTCGCSGGACCGTGGCGCATGTTTCCTGGGAGTTACTGATCATCTTC
 70 90 110
 TTTGAAGAAACATGAAGTTACACTATGTTGCTGTGCTTACTCTAGCCATCCTGATGTTCC
 M K L H Y V A V L T L A I L M F L
 130 150 170
 TGACATGGCTTCCAGAATCACTGAGCTGTAACAAAGCACTCTGTGCTAGTGATGTGAGCA
 T W L P E S L S C N K A L C A S D V S K
 190 210 230
 AATGCCTCATTGAGGAGCTCTGCCAGTGCCGGCGGAGAGGCAATTGCTCCTGCTGTA
 C L I O E L C Q C R P G E G N C S C C K
 250 270 290
 AGGAGTGCATGCTGTGTCTTGGGGCCCTTTGGGACGAGTGCTGTGACTGTGTTGGTATGT
 E C M L C L G A L W D E C C D C V G M C
 310 330 350
 GTAATCCTCGAAATTATAGTGACACACCTCCAACCTTCAAAGAGCACAGTGGAGGAGCTGC
 N P R N Y S D T P P T S K S T V E E L H
 370 390 410
 ATGAACCGATCCCTTCTCTCTTCCGGGCACTCACAGAAGGAGATACTCAGTTGAATTGGA
 E P I P S L F R A L T E G D T Q L N W N
 430 450 470
 ACATCGTTTCTTCCCTGTTGCAGAAGAACTTTCACATCATGAGAATCTGGTTTCATTTT
 I V S F P V A E E L S H H E N L V S F L
 490 510 530
 TAGAACTGTGAACCAGCCACACCACCAAGATGTGTCTGTCCCCAGCAATAATGTTACG
 E T V N Q P H H Q N V S V P S N N V H A
 550 570 590
 CGCCTTATTCAGTGACAAAGAACACATGTGTACTGTGGTTTATTTTGATGACTGCATGT
 P Y S S D K E H M C T V V Y F D D C M S
 610 630 650
 CCATACATCAGTGTAATAATATCCTGTGAGTCCATGGGAGCATCAAATATCGCTGGTTTC
 I H O C K I S C E S M G A S K Y R W F H
 670 690 710
 ATAATGCCTGCTGCGAGTGCATTGGTCCAGAATGTATTGACTATGGTAGTAAACTGTCA
 N A C C E C I G P E C I D Y G S K T V K
 730 750 770
 AATGTATGAACTGCATGTTTTAAAGAAGACAAATGCAAACCAAAGCAACTTAGTAAAATA
 C M N C M F *

FIG 1

10 30 50
 GCACGAGGGCGACTTCGCGGGACCGTGGCGCATGTTTCCTGGGAGTTACTGATCATCTTC
 70 90 110
 TTTGAAGAAACATGAAGTTACACTATGTTGCTGTGCTTACTCTAGCCATCCTGATGTTCC
 M K L H Y V A V L T L A I L M F L
 130 150 170
 TGACATGGCTTCCAGAATCACTGAGCTGTAACAAAGCACTCTGTGCTAGTGATGTGAGCA
 T W L P E S L S C N K A L C A S D V S K
 190 210 230
 AATGCCTCATT CAGGAGCTCTGCCAGTGCCGGCCGGGAGAAGGCAATTGCTCCTGCTGTA
 C L I Q E L C Q C R P G E G N C S C C K
 250 270 290
 AGGAGTGCATGCTGTGTCTTGGGGCCCTTTGGGACGAGTGCTGTGACTGTGTTGGTATGT
 E C M L C L G A L W D E C C D C V G M C
 310 330 350
 GTAATCCTCGAAATTATAGTGACACACCTCCAACCTCAAAGAGCACAGTGGAGGAGCTGC
 N P R N Y S D T P P T S K S T V E E L H
 370 390 410
 ATGAACCGATCCCTTCTCTCTTCCGGGCACTCACAGAAGGAGATACTCAGTTGAATTGGA
 E P I P S L F R A L T E G D T Q L N W N
 430 450 470
 ACATCGTTTCTTTCCCTGTTGCAGAAGAACTTTACATCATGAGAATCTGGTTTCATTTT
 I V S F P V A E E L S H H E N L V S F L
 490 510 530
 TAGAACTGTGAACCAGCCACACCACAGAATGTGTCTGTCCCCAGCAATAATGTTACG
 E T V N Q P H H Q N V S V P S N N V H A
 550 570 590
 CGCCTTATTCCAGTGACAAAGAACACATGTGTACTGTGGTTTATTTTGATGACTGCATGT
 P Y S S D K E H M C T V V Y F D D C M S
 610 630 650
 CCATACATCAGTGTAATAATATCCTGTGAGTCCATGGGAGCATCCAAATATCGCTGGTTTC
 I H Q C K I S C E S M G A S K Y R W F H
 670 690 710
 ATAATGCCTGCTGCGAGTGCATTGGTCCAGAATGTATTGACTATGGTAGTAAACTGTCA
 N A C C E C I G P E C I D Y G S K T V K
 730 750 770
 AATGTATGAACTGCATGTTTTAAAGAAGACAAATGCAAACCAAAGCAACTTAGTAAATA
 C M N C M F *

FIG.1

1	MKLHYVAVLTAILMFLTWLPESLSCNKALCASDVSKCLIQELCQCRPGE	50
	. : : : : :... :: : ... : : : .:	
1	mqllcyfvilfvglapwssl anddgcnvvcgsvvskclitqscqcklnd	50
51	GNCSCCKECLCLGALWDECCDCVGMCPNRYSDTPPTSKSTVEELHEPI	100
	: : . : : :: : .::: :.	
51	..chcckdc nc gelyieccgcl dmcphkhdv ps tprseigdi.egv	97
101	PSLFRALTEGDTQLNWNIVSFPVAEELSHHENLVSFLETVNQPHHQNVSV	150
	. . . : : .::: . .. :..	
98	pe fdt taedde.gwstirfsmragfkqrvqggasgdagn.....	137
151	PSNNVHAPYSSDKEHMCTVVYFDDCMSIHQCKISCESMGASKYRWFHNAC	200
	. .: : . : : .:. : .: : . :	
138	..gngngnagsagvt ctviyvns rankcrqqcesmgassyrwfhdc	185
201	CECIGPECIDYGSKTVKCMNC	221
	: .:. :: .. : ..	
186	cecvgenc nyginesrcrgc	206

FIG.2

DECLARATION FOR PATENT APPLICATION

As a below named inventor, I declare that I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

HUMAN CYTOKINE POLYPEPTIDE

the specification of which [X] is attached hereto or [] was filed on _____ as Application Serial No. _____ and was amended on _____ (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, Section 1.56(a).

I hereby claim foreign priority benefits under 35 U.S.C. § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT international application, which designated at least one country other than the United States listed below, and have also identified below any foreign application for patent or inventor's certificate, or PCT international application having a filing date before that of the application on which priority is claimed. Prior Foreign Application(s):

			<u>Priority</u>	
			<u>Claimed</u>	
			Yes	No
_____ (Number)	_____ (Country)	_____ (Day/Month/Year Filed)		

I hereby claim the benefit under Title 35, United States Code, Section 119(e) of any United States provisional application(s) listed below:

_____ 60/013,613 (Application Serial No.)	_____ March 20, 1996 (Filing Date)
_____ (Application Serial No.)	_____ (Filing Date)

I hereby claim the benefit under Title 35, United States Code, Section 120 of any United States application(s) or under § 365(b) of any PCT international application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, Section 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, Section 1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

_____ 08/820,970 (Application Serial No.)	_____ March 19, 1997 (Filing Date)	_____ Pending (Status: patented, pending, abandoned)
---	--	--

I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith: James H. Davis (Reg. No. 40,582), Kenley K. Hoover (Reg. No. 40,302), Michele M. Wales (Reg. No. 43,975), and Joseph J. Kenny (Reg. No. 43,710) of Human Genome Sciences, Inc., 9410 Key West Avenue, Rockville, Maryland, 20850.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of first joint inventor: Kui Su

Inventor's signature: _____ Date: _____
Residence: 8 Jamaica Way Court, Jamaica Plain, MA 02130 Citizenship: CN
Post Office Address: same as above

Full name of additional joint inventor: Yi Li

Inventor's signature: _____ Date: _____
Residence: 3380 Central Expressway, Santa Clara, CA 95051 Citizenship: CN
Post Office Address: same as above

Full name of additional joint inventor: Haodong Li

Inventor's signature: _____ Date: _____
Residence: 11033 Rutledge Drive, Gaithersburg, MD 20878 Citizenship: CN
Post Office Address: same as above